

Rat MCP-1 ELISA Kit

Catalog Number KRC1011 (96 tests), KRC1012 (2 × 96 tests)

Pub. No. MAN0003978 Rev. 2.0

CAUTION! This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state, and local regulations for disposal.

Note: For safety and biohazard guidelines, see the “Safety” appendix in the *ELISA Technical Guide* (Pub. no. MAN0006706). Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

Product description

The Invitrogen™ Rat MCP-1 ELISA Kit is a solid-phase sandwich Enzyme-Linked Immunosorbent Assay (ELISA). This assay is designed to detect and quantify the level of rat MCP-1 in serum, plasma, buffered solution, or cell culture supernatants. The assay recognizes both natural and recombinant rat MCP-1.

Contents and storage

Upon receipt, store the kit at 2°C to 8°C.

Contents	Cat. No. KRC1011 (96 tests)
Rt MCP-1 Standard, lyophilized; contains 0.1% sodium azide. Refer to vial label for quantity and reconstitution volume	2 vials
Standard Diluent Buffer; contains 0.1% sodium azide	25 mL
Incubation Buffer; contains 0.05% sodium azide	12 mL
Rt MCP-1 Antibody Coated Wells, 96-well strip-well plate	1 plate
Rt MCP-1 Biotin Conjugate; contains 0.1% sodium azide	6 mL
Streptavidin-HRP (100X); contains 3.3 mM thymol	0.125 mL
Streptavidin-HRP Diluent; contains 3.3 mM thymol	25 mL
Wash Buffer Concentrate (25X)	100 mL
Stabilized Chromogen, Tetramethylbenzidine (TMB)	25 mL
Stop Solution	25 mL
Adhesive Plate Covers	3

Materials required but not supplied

- Distilled or deionized water
- Microtiter plate reader with software capable of measurement at or near 450 nm
- Plate washer—automated or manual (squirt bottle, manifold dispenser, or equivalent)
- Calibrated adjustable precision pipettes and glass or plastic tubes for diluting solutions

Before you begin

IMPORTANT! Reagents are lot-specific. Do not mix or interchange different reagent lots from various kit lots.

- Review the **Procedural guidelines** and **Plate washing directions** in the *ELISA Technical Guide* available at thermofisher.com.
- Allow reagents to reach room temperature before use. Mix to redissolve any precipitated salts.

Prepare 1X Wash Buffer

1. Dilute 16 mL of Wash Buffer Concentrate (25X) with 384 mL of deionized or distilled water. Label as 1X Wash Buffer.
2. Store the concentrate and 1X Wash Buffer in the refrigerator. Use the diluted buffer within 14 days.

Sample preparation guidelines

- Refer to the *ELISA Technical Guide* at thermofisher.com for detailed sample preparation procedures.
- Collect samples in pyrogen/endotoxin-free tubes.
- Freeze samples after collection if samples will not be tested immediately. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well (do not vortex) prior to analysis.
- Avoid the use of hemolyzed or lipemic sera. If large amounts of particulate matter are present in the sample, centrifuge or filter sample prior to analysis.

Pre-dilute samples

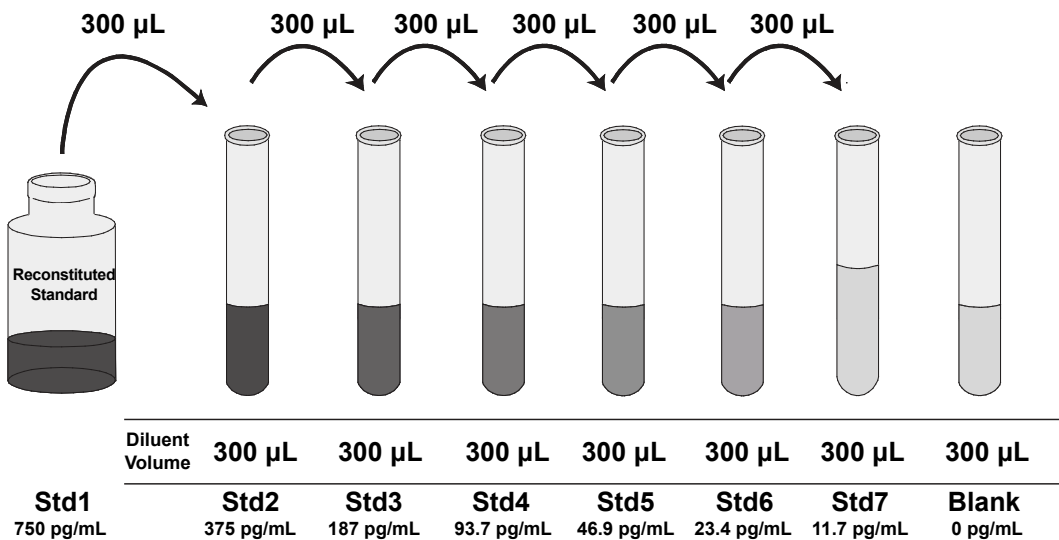
Because conditions may vary, we recommend that each investigator determine the optimal dilution for each application.

- Dilute samples that are >750 pg/mL with Standard Diluent Buffer.

Dilute standards

Note: Use glass or plastic tubes for diluting standards.

1. Reconstitute Rt MCP-1 Standard to 750 pg/mL with Standard Diluent Buffer. Refer to the standard vial label for instructions. Swirl or mix gently and allow the contents to sit for 10 minutes to ensure complete reconstitution. Label as 750 pg/mL rat MCP-1. **Use the standard within 1 hour of reconstitution.**
2. Add 300 μ L Standard Diluent Buffer to each of 7 tubes labeled as follows: 375, 187, 93.7, 46.9, 23.4, 11.7, and 0 pg/mL rat MCP-1.
3. Make serial dilutions of the standard as shown in the following dilution diagram. Mix thoroughly between steps.
4. Discard any remaining reconstituted standard. Return the Standard Diluent Buffer to the refrigerator.



Prepare 1X Streptavidin-HRP solution

Note: Prepare 1X Streptavidin-HRP within 15 minutes of usage.

The Streptavidin-HRP (100X) is in 50% glycerol, which is viscous. To ensure accurate dilution:

1. For each 8-well strip used in the assay, pipet 10 μ L Streptavidin-HRP (100X) solution, wipe the pipette tip with clean absorbent paper to remove any excess solution, and dispense the solution into a tube containing 1 mL of Streptavidin-HRP Diluent. Mix thoroughly.
2. Return the unused Streptavidin-HRP (100X) solution to the refrigerator.

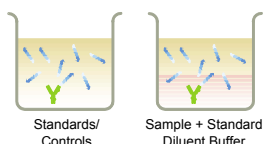
Perform ELISA (Total assay time: 2.5 hours)

IMPORTANT! Perform a standard curve with each assay.

- Allow all components to reach room temperature before use. Mix all liquid reagents prior to use.
- Determine the number of 8-well strips required for the assay. Insert the strips in the frames for use. Re-bag any unused strips and frames, and store at 2°C to 8°C for future use.

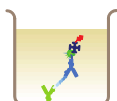


1 Bind antigen



- Add 50 µL of the Incubation Buffer to all wells except the chromogen blanks.
- Add 100 µL of standards or controls to the appropriate wells. Leave the wells for chromogen blanks empty.
- Add 50 µL of **Standard Diluent Buffer** followed by 50 µL of sample (see “Pre-dilute samples” on page 2) to the appropriate wells. Leave the wells for chromogen blanks empty.
- Add 50 µL Rt MCP-1 Biotin Conjugate solution into each well except the chromogen blanks.
- Tap the side of the plate to mix. Cover the plate with a plate cover and incubate for 1 hour and 30 minutes at room temperature.
- Thoroughly aspirate the solution and wash wells 4 times with 1X Wash Buffer.

2 Add Streptavidin-HRP



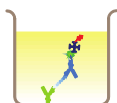
- Add 100 µL 1X Streptavidin-HRP solution (see page 2) into each well except the chromogen blanks.
- Cover the plate with a plate cover and incubate for 30 minutes at room temperature.
- Thoroughly aspirate the solution from the wells and wash wells 4 times with 1X Wash Buffer.

3 Add Stabilized Chromogen



- Add 100 µL Stabilized Chromogen to each well. The substrate solution begins to turn blue.
 - Incubate for 30 minutes at room temperature in the dark.
- Note:** TMB should not touch aluminum foil or other metals.

4 Add Stop Solution



Add 100 µL Stop Solution to each well. Tap the side of the plate to mix. The solution in the wells changes from blue to yellow.

Read the plate and generate the standard curve

1. Read the absorbance at 450 nm. Read the plate within 2 hours after adding the Stop Solution.
2. Use curve-fitting software to generate the standard curve. A 4 parameter algorithm provides the best standard curve fit. Optimally, the background absorbance may be subtracted from all data points, including standards, unknowns and controls, prior to plotting.
3. Read the concentrations for unknown samples and controls from the standard curve. Multiply value(s) obtained for sample(s) by the appropriate factor to correct for the sample dilution.

Note: Dilute samples producing signals greater than that of the highest standard in Standard Diluent Buffer and reanalyze. Multiply the concentration by the appropriate dilution factor.

Performance characteristics

Standard curve example

The following data were obtained for the various standards over the range of 0–750 pg/mL rat MCP-1.

Standard Rat MCP-1 (pg/mL)	Optical Density (450 nm)
750	2.25
375	1.63
187	0.87
93.7	0.44
46.9	0.21
23.4	0.13
11.7	0.09
0	0.06

Inter-assay precision

Samples were assayed 42 times in multiple assays to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	101.6	247.2	381.3
Standard Deviation	9.9	16.6	34.5
% Coefficient of Variation	9.7	6.7	9.0

Intra-assay precision

Samples of known rat MCP-1 concentration were assayed in replicates of 14 to determine precision within an assay.

Parameters	Sample 1	Sample 2	Sample 3
Mean (pg/mL)	110.0	241.6	390.5
Standard Deviation	4.4	13.0	29.6
% Coefficient of Variation	4.0	5.4	7.8

Expected values

A limited number of serum samples (n=5) were assayed with the Rat MCP-1 ELISA Kit. The mean value obtained was 362 pg/mL (range: 253–412 pg/mL).

Sample	Range (pg/mL)	Average (pg/mL)
Serum (n=5)	253–412	362

Linearity of dilution

Rat serum containing 821 pg/mL of measured rat MCP-1 was serially diluted in Standard Diluent Buffer over the range of the assay. Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 0.99.

Sample	Correlation Coefficient
Serum	0.99

Recovery

The recovery of recombinant rat MCP-1 added to rat serum and tissue culture medium containing fetal bovine serum was measured with the Rat MCP-1 ELISA Kit.

Sample	Avg % Recovery
Rat serum	86.0
Tissue culture medium + 1% fetal bovine serum	92.0
Tissue culture medium + 10% bovine serum	101.0

Limited product warranty

Life Technologies Corporation and/or its affiliate(s) warrant their products as set forth in the Life Technologies' General Terms and Conditions of Sale found on Life Technologies' website at www.thermofisher.com/us/en/home/global/terms-and-conditions.html. If you have any questions, please contact Life Technologies at www.thermofisher.com/support.








Sensitivity

The minimum detectable dose of rat MCP-1 is <8 pg/mL. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 30 times, and calculating the corresponding concentration.

Specificity

Buffered solutions of a panel of substances of concentration 10,000 pg/mL were assayed with the Rat MCP-1 ELISA Kit and found to have no cross-reactivity: **human** IL-1 β , IL-3, IL-7, IL-8, IL-15, SCF, RANTES, GM-CSF, TNF- α , IFN- γ , MCP-1; **mouse** IL-2, IL-10, IFN- γ , MCP-1, TNF- α ; **rat** IL-1 β , TNF- α , IFN- γ , MIP-2.

Product label explanation of symbols and warnings

 REF	Catalog Number	 LOT	Batch code		Temperature limitation		Use by		Manufacturer		Consult instructions for use		Caution, consult accompanying documents
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Manufacturer's address: Life Technologies Corporation | 7335 Executive Way | Frederick, MD 21704 | USA

The information in this guide is subject to change without notice.

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