

NuPAGE® Bis-Tris Mini Gels

	Package Contents	<table border="1"> <thead> <tr> <th>Product</th> <th>Quantity</th> </tr> </thead> <tbody> <tr> <td>10% Bis-Tris Gels</td> <td>Box of 2 or 10 gels</td> </tr> <tr> <td>4–12% Bis-Tris Gels</td> <td>Box of 2 or 10 gels</td> </tr> <tr> <td>12% Bis-Tris Gels</td> <td>Box of 2 or 10 gels</td> </tr> </tbody> </table>	Product	Quantity	10% Bis-Tris Gels	Box of 2 or 10 gels	4–12% Bis-Tris Gels	Box of 2 or 10 gels	12% Bis-Tris Gels	Box of 2 or 10 gels
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4–12% Bis-Tris Gels	Box of 2 or 10 gels									
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	Storage Conditions	<ul style="list-style-type: none"> Store at 4–25°C for a 1-year shelf life. Do not freeze. 								
	Required Materials	<ul style="list-style-type: none"> Protein sample and standard NuPAGE® MES or MOPS SDS Running Buffer (20X) NuPAGE® LDS Sample Buffer (4X) NuPAGE® Sample Reducing Agent (10X) NuPAGE® Antioxidant Novex® Power Supply Adapters (Cat. no. ZA10001) if not using a Life Technologies power supply XCell SureLock® Mini-Cell gel running tank 								
	Timing	<p>Run Time: 35 minutes with MES Buffer 50 minutes with MOPS Buffer</p> <p>Voltage: 200 V constant</p>								
	Selection Guide	<p>Protein Gels Go online to view related products.</p>								
	Product Description	<p>NuPAGE® Bis-Tris Gels are precast polyacrylamide gels designed for optimal separation and resolution of small- to medium- sized proteins (1–200 kDa) under denaturing gel electrophoresis conditions.</p> <p>NuPAGE® Bis-Tris Mini Gels are available in the following variations:</p> <ul style="list-style-type: none"> Polyacrylamide percentages: 10%, 4–12%, and 12% Well formats: 1, 9, 10, 12, 15, 17, IPG, and 2D Thicknesses: 1.0 mm and 1.5 mm 								
	Important Guidelines	<ul style="list-style-type: none"> This system is designed for use in the XCell SureLock® Mini-Cell gel running tank. Use the NuPAGE® MES SDS Running Buffer for small proteins or NuPAGE® MOPS SDS Running Buffer for medium-size proteins. 								
	Online Resources	<p>Visit our product page for additional information and protocols. For support, visit www.lifetechnologies.com/support.</p>								



Protocol Outline

- Prepare samples, buffers, and gels.
- Assemble the gel apparatus.
- Load buffer, samples, and standards.
- Perform electrophoresis.

Electrophoresis Protocol

- See page 2 to view a procedure for preparing and running your electrophoresis experiment.

Choosing the Right Gel Type for Your Application

- Review the table in the pop-up to determine the best gel type for your experiment.

Choosing the Right Gel Percentage and Buffer

- Refer to the migration and conversion charts in the pop-up to find the gel best suited for your application. As a general rule, your proteins of interest should migrate through ~70% of the length of the gel for the best resolution. When protein molecular weights are wide ranging or unknown, gradient gels are usually the best choice.

Choosing a Well Format and Gel Thickness

- We offer polyacrylamide gels in a choice of nine well formats and two thicknesses, depending on gel type. When loading large samples (>30 µL), a thicker gel with fewer wells is more appropriate; Bolt™ Bis-Tris Plus gels are the best choice when loading large samples. When blotting, however, proteins will transfer more easily from a thinner gel.

Choosing a Protein Standard for your Application

Choose a Life Technologies™ standard based on your experiment:

Pre-Stained: SeeBlue® Plus2 Pre-Stained Standard or Novex® Sharp Pre-Stained Protein Standard

Unstained: Novex® Sharp Unstained Protein Standard or Mark12™ Unstained Standard

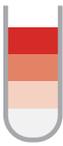
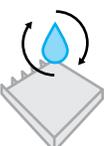
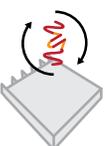
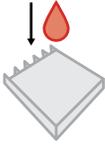
Western: MagicMark™ XP Western Protein Standard

For all other specialty standards, please view further information [here](#).

Limited Product Warranty and Disclaimer Details

NuPAGE® Bis-Tris Mini Gel Electrophoresis Protocol

Follow the procedure below to prepare for and perform SDS polyacrylamide gel electrophoresis using NuPAGE® Bis-Tris Mini Gels.

Timeline	Steps	Procedure Details																		
	Prepare samples	<table border="1"> <thead> <tr> <th>Components</th> <th>Reduced Sample</th> <th>Non-Reduced Sample</th> </tr> </thead> <tbody> <tr> <td>Sample</td> <td>x μL</td> <td>x μL</td> </tr> <tr> <td>NuPAGE® LDS Sample Buffer (4X)</td> <td>2.5 μL</td> <td>2.5 μL</td> </tr> <tr> <td>NuPAGE® Reducing Agent (10X)</td> <td>1 μL</td> <td>--</td> </tr> <tr> <td>Deionized Water</td> <td>to 6.5 μL</td> <td>to 7.5 μL</td> </tr> <tr> <td>Total Volume</td> <td>10 μL</td> <td>10 μL</td> </tr> </tbody> </table> <p>Heat samples at 70°C for 10 minutes. Prepare 1X Sample Buffer for dilutions of samples, if needed.</p>	Components	Reduced Sample	Non-Reduced Sample	Sample	x μL	x μL	NuPAGE® LDS Sample Buffer (4X)	2.5 μL	2.5 μL	NuPAGE® Reducing Agent (10X)	1 μL	--	Deionized Water	to 6.5 μL	to 7.5 μL	Total Volume	10 μL	10 μL
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Total Volume	10 μL	10 μL																		
	Prepare buffers	<p>Add 50 mL of 20X NuPAGE® MES or MOPS SDS Running Buffer to 950 mL of deionized water to prepare 1X SDS Running Buffer.</p> <p>For reduced samples, prepare the running buffer for the Upper Buffer Chamber by adding 500 μL of NuPAGE® Antioxidant to 200 mL 1X SDS Running Buffer.</p>																		
	Prepare gels	<ol style="list-style-type: none"> Remove the comb, and rinse the gel wells three times using 1X Running Buffer. Remove the white tape near the bottom of the gel cassettes. Place the gels in the XCell SureLock® Mini-Cell gel running tank. Fill the gel wells with 1X Running Buffer. 																		
	Load gels	<p>Load the appropriate volume and protein mass of your sample on the gel. Then, load your standards.</p>																		
	Load buffers	<p>Fill the Upper (200 mL) and Lower (600 mL) Buffer Chambers with the appropriate 1X Running Buffer.</p>																		
	Run	<p>Note: If you are not using a Life Technologies™ power supply, install the Novex® Power Supply Adapters (Catalog number ZA10001).</p> <p>When using MES Running Buffer, run for 35 minutes at 200 V constant. When using MOPS Running Buffer, run for 50 minutes at 200 V constant.</p>																		