

Dynabeads® Untouched™ Human T Cells

Catalog no. 11344D

Store at 2°C to 8°C

Rev. Date: February 2012 (Rev. 002)

Kit Contents

Kit contents	Volume
Depletion Dynabeads®	2 × 5 mL
Antibody Mix (Human T Cells)	2 mL

Kit capacity
PBMC: ~1 × 10⁹

Depletion Dynabeads® contains 4×10^8 beads/mL in phosphate buffered saline (PBS), pH 7.4, with 0.1% bovine serum albumin (BSA) and 0.02% sodium azide as a preservative. Antibody Mix contains monoclonal mouse anti-human IgG antibodies in PBS with 0.5% BSA and 0.02% sodium azide.

Caution: Sodium azide may react with lead and copper plumbing to form highly explosive metal azides.

Product Description

This product is intended for isolation of untouched human T cells from peripheral blood mononuclear cells (PBMC) by depleting B cells, NK cells, monocytes, platelets, dendritic cells, granulocytes and erythrocytes. Isolated T cells are bead- and antibody-free and are suitable for any downstream application (fig. 1).

A mixture of mouse IgG antibodies against the non-T cells is added to the starting sample. Depletion Dynabeads® are added and bind to the antibody-labeled cells during a short incubation. The bead-bound cells are subsequently separated on a magnet and discarded. The supernatant contains the untouched human T cells.

Downstream Applications

Isolated T cells can be used in any downstream application e.g. flow cytometry, T cell culture, T cell activation and expansion, functional assays, molecular studies.

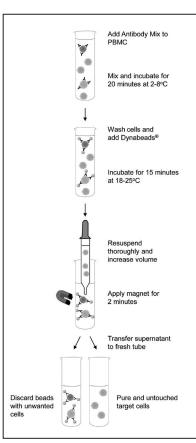


Figure 1: Isolation principle for untouched T Cells.

Required Materials

- Magnet (DynaMag[™]) See www.lifetechnologies.com/magnets for recommendations.
- Mixing device with tilting and rotation, e.g. HulaMixer® Sample Mixer.
- Heat inactivated Fetal Bovine Serum (FBS)/Fetal Calf Serum (FCS).
- Isolation Buffer: PBS (Ca²⁺ and Mg²⁺ free) supplemented with 0.1% BSA and 2 mM EDTA.

Note: BSA can be replaced by human serum albumin (HSA) or 2% FBS/FCS. EDTA can be replaced by 0.6% sodium citrate.

 Lymphoprep[®] for PBMC preparation (Axis Shield PoC, Norway, www.axis-shield-poc.com).

General Guidelines

- Visit www.lifetechnologies.com/samplepreparation for recommended sample preparation procedures.
- Use a mixer that provides tilting and rotation of the tubes to ensure that Dynabeads[®] do not settle in the tube.
- This product should not be used with the MPC[™]-1 magnet (Cat. no. 12001D).
- Follow the recommended volumes and incubation times.
- Avoid air bubbles (foaming) during pipetting.
- Keep the buffers cold.

Protocol

Wash Dynabeads®

See Table 1 for volume recommendations.

- 1. Resuspend the Dynabeads® in the vial (i.e vortex for >30 sec, or tilt and rotate for 5 min).
- 2. Transfer the desired volume of Dynabeads® to a tube.
- 3. Add the same volume of Isolation Buffer, or at least 1 mL, and resuspend.
- 4. Place the tube in a magnet for 1 min and discard the supernatant.
- 5. Remove the tube from the magnet and resuspend the washed Dynabeads® in the same volume of Isolation Buffer as the initial volume of Dynabeads® (step 2).

Prepare Cells

Prepare a PBMC suspension according to "General Guidelines". Resuspend the cells at 1×10^8 cells/mL in Isolation Buffer.

Isolation Procedure

This protocol is based on 5×10^7 PBMC, but is directly scalable from 1×10^7 to 5×10^8 cells, according to Table 1.

- 1. Transfer 500 μ L (5 × 10⁷) PBMC in Isolation Buffer to a tube.
- 2. Add 100 µL heat inactivated FBS/FCS.
- 3. Add 100 µL of Antibody Mix.
- 4. Mix well and incubate for 20 min at 2°C to 8°C.
- 5. Wash the cells by adding 4 mL Isolation Buffer. Mix well by tilting the tube several times and centrifuge at $350 \times g$ for 8 min at 2°C to 8°C. Discard the supernatant.
- 6. Resuspend the cells in 500 µL Isolation Buffer.
- 7. Add 500 µL pre-washed Dynabeads®.
- 8. Incubate for 15 min at 18°C to 25°C with gentle tilting and rotation.
- 9. Add 4 mL Isolation Buffer. (When working with lower cell volumes, never use less than 1 mL Isolation Buffer).
- 10. Resuspend the bead-bound cells thoroughly by pipetting >10 times using a pipette with a narrow tip opening. Avoid foaming.
- 11. Place the tube in the magnet for 2 min. Transfer the supernatant containing the untouched human T cells, to a new larger tube.
- 12. Add 4 mL Isolation Buffer to the tube containing the Dynabeads® and resuspend the bead-bound cells by pipetting as described in step 10.
- 13. Place the tube in the magnet for 2 min.
- 14. Combine the two supernatants.

Table 1: Volumes for isolation of human T cells. This protocol is scalable from 1×10^7 to 5×10^8 PBMC.

Step	Step description	Volumes per 5 × 10 ⁷ PBMC	Volumes per 2 × 10 ⁸ PBMC
	Recommended tube	5–7 mL tubes	15 mL tubes
	Recommended magnet	DynaMag [™] -5	DynaMag [™] -15
1	Cell volume	500 μL	2 mL
2	FBS/FCS	100 μL	400 μL
3	Antibody Mix	100 μL	400 μL
5*	Wash cells (Isolation Buffer)	~4 mL	~10 mL
6	Resuspend cells (Isolation Buffer)	500 μL	2 mL
7**	Depletion Dynabeads®	500 μL	2 mL
9-12*	Increase volume (Isolation Buffer)	2 × ~4 mL	2 × ~10 mL

^{*} Adjust the Isolation Buffer volumes to fit to the tube you are using.

Description of Materials

Depletion Dynabeads® are uniform, superparamagnetic polymer beads (4,5 μ m diameter) coated with a monoclonal human anti-mouse IgG antibody. The antibody coated onto Dynabeads® recognizes all mouse IgG subclasses and is Fc-specific. The Antibody Mix contains mouse IgG antibodies for CD14, CD16 (specific for CD16a and CD16b), CD19, CD36, CD56, CDw123 and CD235a (Glycophorin A). Supplied in PBS with 0.5% BSA and 0.02% sodium azide.

Related Products

Product	Cat. no.
DynaMag [™] -5	12303D
DynaMag [™] -15	12301D
DynaMag [™] -50	12302D
HulaMixer® Sample Mixer	15920D
Phosphate Buffered Saline	10010-023

REF on labels is the symbol for catalog number.

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^{**} When incubating, tilt and rotate so the cells and beads are kept in the bottom of the tube.

Do not perform end-over-end mixing if the volume is small relative to the tube size.