gibco *Life* technologies CHO-S-SFM II

Description

CHO-S-SFM II, is a complete, serum-free, low-protein (<100 μ g/ml) medium optimized for the growth and maintenance of Chinese Hamster Ovary (CHO) cells in suspension culture and for the production of recombinant proteins expressed by CHO cells. This ready-touse medium is suitable for the suspension culture of CHO and other cells in batch, continuous, and perfusion culture systems. Component deficient CHO-S-SFM II, is formulated without hypoxanthine and thymidine for use in dihydrofolate reductase (DHFR) amplified systems.

Product	Catalog no.	Amount	Storage	Shelf life*
CHO-S-SFM II	12052-114 12052-098	500 mL 1000 mL	2°C to 8°C; Protect from light 2°C to 8°C; Protect from light	12 months 12 months
CHO-S-SFM II, powder	22052-021 22052-047	10 × 1 L 1 × 50 L	2°C to 8°C; Store dark and dry	18 months
CHO-S-SFM II (without hypoxanthine and thymidine)	31033-020	500 mL	2°C to 8°C; Protect from light	12 months

* Shelf life duration is determined from Date of Manufacture.

Product use

For Research Use Only. Not for use in diagnostic procedures.

Product use (Cat. no. 12052 only)

Caution: For manufacturing, processing, or repacking.

Important information

CHO-S-SFM II is a complete medium, formulated with L-glutamine, and requires no further supplementation. Addition of hypoxanthine•Na at 1.66 mg/L and thymidine at 0.28 mg/L to component deficient CHO-S-SFM II, reconstitutes the original formulation of CHO-S-SFM II.

Safety information

Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

Human origin materials are non-reactive (donor level) for anti-HIV 1 & 2, anti-HCV and HBsAg. Handle in accordance with established bio-safety practices.

Prepare media

Reconstitution

- 1. Add CHO-S-SFM II powder to 950 mL room temperature cell culture grade distilled water. Rinse the inside of package to remove all traces of powder.
- 2. Mix with gentle stirring until medium dissolves completely. **Do not heat**.
- 3. Add 2.45 g Sodium Bicarbonate (NaHCO₃) per liter of medium. Stir until dissolved.
- 4. Adjust medium to pH 8.0 with 1N NaOH while stirring. Slowly bring the pH to 7.0–7.1 with 1N HCl (Filtration will cause the pH to rise 0.1–0.3 units; final post-filtration pH should be 7.2–7.4).
- Add deionized or distilled water to final volume of 1000 mL. Check pH and osmolality (Osmolality should be 320–345 mOsm/kg).
- Filter sterilize through a 0.2-µm low extractables, low binding filter into clean, sterile containers. Store at 2°C to 8°C protected from light until use.

Culture conditions

Media: CHO-S-SFM II

Cell line: Chinese Hamster Ovary (CHO)

Culture type: Supension

Culture vessels: shake flask or spinner bottle or bioreactor system. **Temperature range:** 36°C to 38°C

Incubator atmosphere: Humidified atmosphere of 8% CO₂ in air. Ensure proper gas exchange and minimize exposure of cultures to light.

Adapt CHO Cells to CHO-S-SFM II

Sequential adaptation of CHO cells from serum supplemented (or from the original CHO-S-SFM) may be required.

Note: It is critical that cell viability be at least 90% and cells be in the mid-logarithmic phase of growth prior to adaptation.

- 1. Subculture CHO cells grown in conventional medium with 5–10% serum or other serum-free medium into a 50:50 ratio of CHO-S-SFM II to the original media. During the adaptation procedure use a seeding density of 3×10^{5} – 5×10^{5} viable cells/mL.
- 2. Subculture cells when viable cell density reaches of 1×10^6 cells/mL.
- 3. Once consistent cell growth has been achieved, passage cells stepwise increasing the ratio of complete CHO-S-SFM II to original medium (75:25 followed by 90:10) until the cells are transferred into 100% CHO-S-SFM II. Multiple passages at each step may be required.
- 4. After several passages in 100% CHO-S-SFM II, the viable cell count should reach 1×10^{6} - 3×10^{6} cells/mL with viability exceeding 85% within 4–5 days of passage. At this stage the culture is considered to be adapted.

Note: It is not advisable to attempt to adapt cells already growing in serum-free formulations other than Gibco CHO-S-SFM to Gibco CHO-S-SFM II. Adaptation of cells grown in different serum-free media may be affected by selection of subpopulation(s) to specific components.

Cryopreservation

Prepare the desired quantity of cells in a tissue culture flask, harvesting in mid-log phase of growth with viability >90%. Reserve the conditioned medium to prepare cryopreservation medium.

- 1. Determine the viable cell density using a Countess[®] Automated Cell Counter, or alternative automated or manual method, and calculate the required volume of cryopreservation medium to give a final cell density of 0.5×10^7 -1 $\times 10^7$ cells/mL.
- Prepare the required volume of cryopreservation medium of 92.5% CHO-S-SFM II (50:50 ratio of fresh to conditioned media) +7.5% DMSO and store at 4°C until use.

Important: Prepare cryopreservation medium on the day of intended use.

- 3. Harvest cells by centrifugation at $100 \times g$ for 5–10 minutes. Resuspend the cell pellet in the pre-determined volume of 4°C cryopreservation medium.
- 4. Dispense aliquots of this suspension into cryovials according to the manufacturer's specifications (i.e., 1.5 mL in a 2-mL cryovial).
- 5. Achieve cryopreservation in an automated or manual controlled rate freezing apparatus following standard procedures (1°C decrease per minute).
- 6. Transfer frozen cells to liquid nitrogen, (vapor phase) storage at -200°C to -125°C is recommended.

Note: Check viability of cryopreserved cells 24 hours after storage of vials in liquid nitrogen. See **Recovery.**

Recovery

- 1. Rapidly thaw (<1 minute) frozen vial of cells in a 37°C water bath.
- 2. Transfer the entire contents of the cryovial into a 125-mL shake flask containing 28.5 mL of pre-warmed CHO-S-SFM II.
- 3. Incubate at 37° C in a humidified atmosphere of 8% CO₂ in air on an orbital shaker platform rotating at 115–135 rpm. Loosen flask caps to allow for gas exchange.
- 4. Subculture cells in mid-logarithmic phase 3–5 days postthaw at a seeding density of 3×10^5 viable cells/mL. Subculture cells a minimum of 3 passages before use in other applications.

Note: Do not centrifuge CHO cells as they are extremely fragile upon recovery from cryopreservation.

Related products

Product	Catalog no.
HT Supplement, (100X), liquid	11067
Anti-Clumping Agent	0010057
CHO CD EfficientFeed [™] Kit	A10241
CHO CD EfficientFeed [™] B AGT [™]	A12456
CD EfficientFeed TM C AGT TM	A13275
CHO-S [™] Cells (cGMP banked) and Media Kit	A11557-01
Trypan Blue Stain	15250
Countess [®] Automated Cell Counter	C10227

Explanation of symbols and warnings

The symbols present on the product label are explained below:

X		LOT				REF
Temperature Limitation	Manufacturer	Batch code		Use By:		Catalog number
<u>_</u> !	i		**		STERILE A	
Caution, consult accompanying documents	Consult instructions for use		Keep away from light		Sterilized using aseptic processing techniques	

Limited Product Warranty

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