

CD62L antibodies, mouse

For research use only

9 µg equal 60 tests, 30 µg equal 200 tests. One test corresponds to labeling of 10⁶ cells.

Product	Content	Order no.
CD62L-FITC ¹	30 µg in 1 mL	130-102-465
CD62L-PE	9 µg in 300 µL	130-102-907
CD62L-PE	30 µg in 1 mL	130-102-543
CD62L-APC	9 µg in 300 µL	130-102-931
CD62L-APC	30 µg in 1 mL	130-102-453
CD62L-VioBlue	9 µg in 300 µL	130-103-135
CD62L-VioBlue	30 µg in 1 mL	130-102-425
CD62L-PE-Vio770	9 µg in 300 µL	130-104-520
CD62L-PE-Vio770	30 µg in 1 mL	130-104-471
CD62L-PerCP-Vio700	9 µg in 300 µL	130-107-072
CD62L-PerCP-Vio700	30 µg in 1 mL	130-107-046
CD62L-Biotin	9 µg in 300 µL	130-102-007
CD62L-Biotin	30 µg in 1 mL	130-101-933

¹Not recommended for cells that are labeled with MACS MicroBeads using the same antigen.

Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

Technical data and background information

Antigen	CD62L
Clone	MEL14-H2.100
Isotype	rat IgG2ak
Isotype control	Rat IgG2a – isotype control antibodies
Alternative names of antigen	Sell, L-selectin, LECAM-1, Lnhp, Ly-22, Ly-m22, Lyam-1, Lyam1, LAM-1
Molecular mass of antigen [kDa]	38
Distribution of antigen	B cells, bone marrow, eosinophils, granulocytes, Langerhans cells, lymphocytes, monocytes, neutrophils, NK cells, red blood cells, spleen, T cells, thymocytes
Product format	Antibodies are supplied in buffer containing stabilizer and 0.05% sodium azide.
Storage	Store protected from light at 2–8 °C. Do not freeze.

Clone MEL14-H2.100 recognizes mouse CD62L (L-selectin), a member of the selectin family of cell adhesion molecules. CD62L is important for the homing of lymphocytes to peripheral lymph nodes through high endothelial venules. It also contributes to leukocyte emigration into acute inflammatory sites. CD62L is a glycoprotein expressed on the cell surface of most thymocytes, T cells, B cells, monocytes, dendritic cells, neutrophils, and eosinophils. It is highly expressed on naive T cells and down-regulated upon differentiation due to activation-dependent shedding. CD62L is also expressed on a subset of CCR7^{high}CD44^{high} central memory T cells and on CD4⁺CD25⁺ regulatory T cells.

Reagent requirements

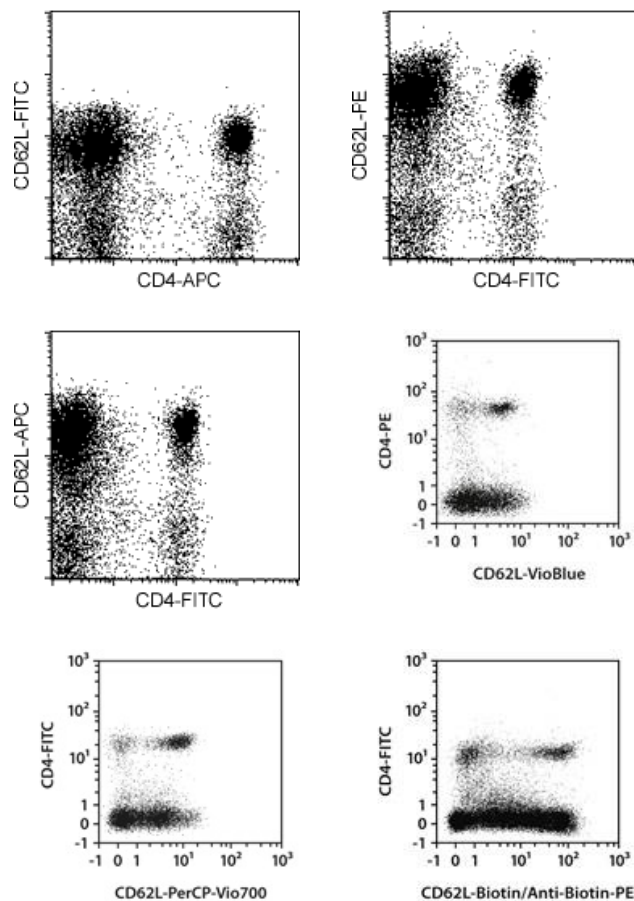
- Buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS[®] BSA Stock Solution (# 130-091-376) 1:20 with autoMACS[®] Rinsing Solution (# 130-091-222). Keep buffer cold (2–8 °C).
Note: EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). Buffers or media containing Ca²⁺ or Mg²⁺ are not recommended for use.
- (Optional) FcR Blocking Reagent, mouse (# 130-092-575) to avoid Fc receptor-mediated antibody labeling.
- (Optional) Fluorochrome-conjugated anti-biotin antibodies, e.g., Anti-Biotin-PE (# 130-090-756) as secondary antibody reagent in combination with biotinylated antibodies.
- (Optional) Propidium Iodide Solution (# 130-093-233) for flow cytometric exclusion of dead cells without fixation.
- (Optional) Fixation and Dead Cell Discrimination Kit (# 130-091-163) for cell fixation and flow cytometric exclusion of dead cells.

Protocol for cell surface staining

- The recommended antibody dilution for labeling of cells and subsequent analysis by flow cytometry is 1:10 for up to 10⁶ cells/50 µL of buffer.
 - Volumes given below are for up to 10⁶ nucleated cells. When working with fewer than 10⁶ cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes accordingly (e.g. for 2×10⁶ nucleated cells, use twice the volume of all indicated reagent volumes and total volumes).
1. Determine cell number.
 2. Centrifuge cell suspension at 300×g for 10 minutes. Aspirate supernatant completely.
 3. Resuspend up to 10⁶ nucleated cells per 45 µL of buffer.
 4. Add 5 µL of the antibody.
 5. Mix well and incubate for 10 minutes in the dark in the refrigerator (2–8 °C).
Note: Higher temperatures and/or longer incubation times may lead to non-specific cell labeling. Working on ice requires increased incubation times.
 6. Wash cells by adding 1–2 mL of buffer and centrifuge at 300×g for 10 minutes. Aspirate supernatant completely.
 7. (Optional) If biotinylated antibody was used, resuspend the cell pellet in 100 µL of buffer, add 10 µL of fluorochrome-conjugated anti-biotin antibody, and continue as described in steps 5 and 6.
 8. Resuspend cell pellet in a suitable amount of buffer for analysis by flow cytometry or fluorescence microscopy.

Examples of immunofluorescent staining

Mouse splenocytes were stained with CD62L antibodies as well as with CD4 antibodies, and analyzed by flow cytometry. Cell debris and dead cells were excluded from the analysis based on scatter signals and propidium iodide fluorescence.



Warranty

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