

# CD4 antibodies, mouse

## For research use only

9 μg equal 60 tests, 30 μg equal 200 tests. One test corresponds to labeling of 10<sup>6</sup> cells.

Content	Order no.
9 μg in 300 μL	130-102-779
30 μg in 1 mL	130-102-541
9 μg in 300 μL	130-102-780
30 μg in 1 mL	130-102-619
9 μg in 300 μL	130-102-782
30 μg in 1 mL	130-102-597
9 μg in 300 μL	130-102-774
30 μg in 1 mL	130-102-456
9 μg in 300 μL	130-102-777
30 μg in 1 mL	130-102-444
9 μg in 300 μL	130-102-784
30 μg in 1 mL	130-102-411
9 μg in 300 μL	130-102-786
30 μg in 1 mL	130-102-179
30 μg in 1 mL	130-102-271
9 μg in 300 μL	130-102-024
30 μg in 1 mL	130-101-962
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<sup>&</sup>lt;sup>1</sup>Not recommended for cells that are labeled with MACS MicroBeads using the same antigen.

### **Warnings**

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

### Technical data and background information

Antigen CD4
Clone GK1.5
Isotype rat IgG2bk

**Isotype control** Rat IgG2b – isotype control antibodies

Alternative names of antigen L3T4, Ly-4, Leu-3, T4

Molecular mass of antigen [kDa] 48

**Distribution of antigen** dendritic cells, granulocytes, Langerhans cells, lymphocytes,

macrophages, monocytes, neutrophils, T cells, T helper cells,

thymocytes

**Product format** Antibodies are supplied in buffer containing stabilizer and 0.05%

sodium azide.

**Fixation** With the exception of the VioGreen conjugate the antibody is

suited for staining of formaldehyde-fixed cells.

**Storage** Store protected from light at 2–8 °C. Do not freeze.

Clone GK1.5 recognizes the mouse CD4 (L3T4) antigen. In mice, CD4 is expressed on T helper cells, regulatory T cells, and at lower levels on subpopulations of NKT cells and dendritic cells. It is furthermore detected on most thymocytes (CD4<sup>+</sup>CD8<sup>+</sup> and CD4<sup>+</sup>CD8<sup>-</sup> thymocytes).

### Reagent requirements

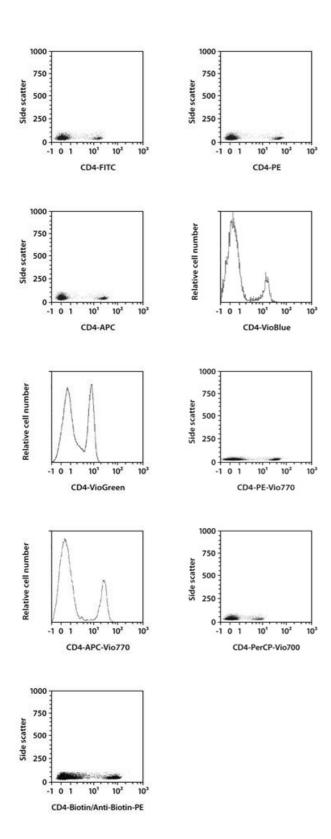
- Buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS<sup>®</sup> BSA Stock Solution (# 130-091-376) 1:20 with autoMACS<sup>®</sup> Rinsing Solution (# 130-091-222). Keep buffer cold (2-8 °C). Note: EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). Buffers or media containing Ca<sup>2+</sup> or Mg<sup>2+</sup> are not recommended for use.
- (Optional) FcR Blocking Reagent, mouse (# 130-092-575) to avoid Fc receptor-mediated antibody labeling.
- (Optional) Fluorochrome-conjugated anti-biotin antibodies, e.g., Anti-Biotin-PE (# 130-090-756) as secondary antibody reagent in combination with biotinylated antibodies.
- (Optional) Propidium Iodide Solution (# 130-093-233) for flow cytometric exclusion of dead cells without fixation.
- (Optional) Fixation and Dead Cell Discrimination Kit (# 130-091-163) for cell fixation and flow cytometric exclusion of dead cells.

### Protocol for cell surface staining

- The recommended antibody dilution for labeling of cells and subsequent analysis by flow cytometry is 1:10 for up to 10<sup>6</sup> cells/50 µL of buffer.
- Volumes given below are for up to 10<sup>6</sup> nucleated cells. When working with fewer than 10<sup>6</sup> cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes accordingly (e.g. for 2×10<sup>6</sup> nucleated cells, use twice the volume of all indicated reagent volumes and total volumes).
- 1. Determine cell number.
- 2. Centrifuge cell suspension at 300×g for 10 minutes. Aspirate supernatant completely.
- 3. Resuspend up to  $10^6$  nucleated cells per 45  $\mu$ L of buffer.
- 4. Add 5 µL of the antibody.
- Mix well and incubate for 10 minutes in the dark in the refrigerator (2-8 °C).
   Note: Higher temperatures and/or longer incubation times may lead to non-specific cell labeling.
   Working on ice requires increased incubation times.
- 6. Wash cells by adding 1–2 mL of buffer and centrifuge at 300×g for 10 minutes. Aspirate supernatant completely.
- 7. (Optional) If biotinylated antibody was used, resuspend the cell pellet in 100  $\mu$ L of buffer, add 10  $\mu$ L of fluorochrome-conjugated anti-biotin antibody, and continue as described in steps 5 and 6.
- 8. Resuspend cell pellet in a suitable amount of buffer for analysis by flow cytometry or fluorescence microscopy.

### **Examples of immunofluorescent staining**

Mouse splenocytes were stained with CD4 antibodies and analyzed by flow cytometry. Cells stained with CD4-Biotin were stained with Anti-Biotin-PE in addition. Cell debris and dead cells were excluded from the analysis based on scatter signals and propidium iodide fluorescence or 4',6-diamidino-2-phenylindole (DAPI) fluorescence, as in the case of CD4-PE-Vio770, CD4-APC-Vio770, CD4-PerCP-Vio700.



### Warranty

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