# Essential 8<sup>™</sup> Adaptation Kit

# Description

rhLaminin-521 is a recombinant human protein that provides a defined surface for feeder-free culture of human pluripotent stem cells (PSCs). rhLaminin-521 provides optimal PSC survival following feeder-dependent to feeder-free transitions when used in conjunction with Essential 8<sup>™</sup> Medium.

Product*	Catalog no.	Amount	Storage	Shelf life
Essential 8 <sup>™</sup> Adaptation Kit contains:	A25935	1 Kit	See below	See below
Essential 8™ Basal Medium Essential 8™ Supplement (50X)	A1517001	500 mL 10 mL	2°C to 8°C. Protect from light. –20°C to –5°C. Protect from light.	1 year**
rhLaminin-521	A29248	100 µg	-30°C to -10°C	2 years***

\*Essential 8<sup>™</sup> Medium (Cat. no. A1517001) and rhLaminin-521 (Cat. no. A29248) are also sold separately.

\*\* Shelf life duration is determined from Date of Manufacture when stored at recommended storage condition.

\*\*\* Shelf life duration is determined from Date of Receipt when stored at recommended storage condition.

## **Product use**

For Research Use Only. Not for use in diagnostic procedures.

## Important information

- Thaw rhLaminin-521 slowly at 2°C to 8°C. Avoid extended exposure of protein to ambient temperatures. For long coating procedures the laminin stock solution should be kept on ice.
- Once thawed, rhLaminin-521 stock is stable for up to 3 months when stored at 2°C to 8°C.
- Divide thawed rhLaminin-521 into usage-size aliquots and store in a non-frost-free freezer at -30°C to -10°C. Avoid repeated freeze-thaw cycles.
- Plates can be coated in advance of experiments, parafilm sealed, and stored at 2°C to 8°C under aseptic conditions for up to 2 weeks. Do not allow the culture surface to dry.

# Safety information

Read the Safety Data Sheets (SDSs) and follow the handling instructions. Wear appropriate protective eyewear, clothing, and gloves.

# **Culture conditions**

Culture type: Adherent feeder-free

Substrate: rhLaminin-521

Diluent: DPBS, calcium, magnesium (Cat. no. 14040)

Recommended media: Essential 8<sup>™</sup> Medium (Cat. no. A1517001)

**Recommended passaging reagent:** TrypLE<sup>™</sup> Select (Cat. no. 12563) **Temperature range:** 36°C to 38°C

**Incubator atmosphere:** Humidified atmosphere of 5% CO<sub>2</sub>. Ensure that proper gas exchange is achieved in culture vessels.

# Working concentration

The optimal working concentration of rhLaminin-521 is cell line dependent and must be determined empirically. We recommend using an initial coating concentration of  $0.5 \,\mu\text{g/cm}^2$  on the culture surface. Prior to coating culture vessels, calculate the working concentration according to the formula below and dilute the stock appropriately. Refer to Table 1 for culture surface area and required coating volumes.

Working conc. = Coating conc.  $\times \frac{Culture \ surface \ area}{Vol. required \ for \ surface \ area}$ 

$$Dilution factor = \frac{Stock \ concentration \ (100 \ \frac{\mu g}{mL})}{Working \ concentration}$$

For example, to coat a 60-mm dish at a coating concentration of  $0.5 \ \mu\text{g/cm}^2$ , you will need to prepare 4 mL of diluted rhLaminin-521 solution (20 cm<sup>2</sup>/dish surface area and 4 mL of diluted rhLaminin-521/dish; see Table 1) at the following working concentration:

Working conc. = 
$$0.5 \frac{\mu g}{cm^2} \times \frac{20 cm^2}{4 mL} = 2.5 \frac{\mu g}{mL}$$

$$Dilution \ factor \ = \frac{100 \ \mu g/mL}{2.5 \ \mu g/mL} = 40X \ (i.e., 1:40 \ dilution)$$

# Coat culture vessels with rhLaminin-521

Instructions for coating a 60-mm culture dish with rhLaminin-521 at a coating concentration of  $0.5 \,\mu\text{g/cm}^2$  are provided below. For volumes used in other culture vessels, refer to Table 1. To calculate the working concentration of rhLaminin-521 used with other coating concentrations and to determine the appropriate dilution factor, use the equations above.

- 1. Upon receipt, thaw the vial of rhLaminin-521 slowly at 2°C to 8°C, mix by gentle trituration, and prepare usage size aliquots in polypropylene tubes. Freeze aliquots at -30°C to -10°C or store aliquots at 2°C to 8°C for up to 3 months.
- To coat a 60-mm dish, add 100 μL aliquot of rhLaminin-521 into a 15-mL conical tube containing 4 mL of sterile DPBS containing calcium and magnesium (Cat. no. 14040). Gently resuspend by pipetting the rhLaminin-521 dilution up and down. Note: This results in a working concentration of 2.5 μg/mL (i.e., a 1:40 dilution).
- Add the diluted rhLaminin-521 solution to the 60-mm dish (refer to Table 1 for the recommended volumes for other culture vessels). When used to coat a 60-mm dish (20 cm<sup>2</sup>) at 4 mL/well, the final coating concentration will be 0.5 µg/cm<sup>2</sup>.
- Incubate the plates in a 37°C, 5% CO<sub>2</sub> incubator for 2 hours for efficient coating.
   Note: Alternatively, the plate can be coated at 2°C to 8°C overnight. Do not allow the culture vessel to dry. Prior to use, pre-warm the culture vessel to room temperature for at least 1 hour.
- 5. Immediately prior to plating of cells, aspirate the rhLaminin-521 solution and discard. It is not necessary to rinse off the culture vessel after the removal of rhLaminin-521. Cells can be passaged directly onto the rhLaminin-521-coated culture vessels.

#### Adapt cells to feeder-free culture in Essential 8™ Medium

Follow the instructions below to adapt feeder-dependent PSC cultures to feeder-free conditions in Essential 8<sup>™</sup> Medium on rhLaminin-521-coated culture vessels. The volumes given in the procedure are for 60-mm culture dishes. For volumes used in other culture vessels, refer to Table 2.

- 1. When the feeder-dependent cultures reach passaging confluency (60%–85% confluent with round colonies that are not overcrowded), the cells are ready for adaptation to feeder-free culture conditions.
- 2. Coat culture vessels with rhLaminin-521 per instructions noted above.
- 3. Prepare a 1 mg/mL Collagenase Type IV solution in DMEM/F12 with GlutaMAX<sup>™</sup> Supplement and filter sterilize using a 0.2-um filter unit.
- 4. Aspirate the spent medium from the culture vessel.
- 5. Rinse the vessel once with 4 mL of Dulbecco's Phosphate Buffered Saline (DPBS) without calcium or magnesium.
- 6. Add 2 mL of 1 mg/mL Collagenase Type IV, pre-warmed to 37°C.
- Incubate the vessel for ~45 minutes in a 37°C, 5% CO<sub>2</sub> incubator. Note: Stop the incubation when the edges of the colonies begin to curl from the plate. Do not over-incubate.
- Add 2 mL of Essential 8<sup>™</sup> Medium and gently dislodge the colonies from the plate by washing off colonies with a 5-mL serological pipette. Repeat trituration until the desired cluster size is achieved.
- 9. Transfer the suspended colony clusters into a 15-mL conical tube.
- 10. Add 2 mL of Essential 8<sup>™</sup> Medium to dislodge the remaining colonies and transfer them to the 15-mL conical tube.
- 11. Let the colony fragments sediment to the bottom of the 15-mL conical tube for 5 minutes by gravity.
- 12. Discard the supernatant, add 4 mL of Essential 8<sup>™</sup> Medium, and gently resuspend the sedimented colony fragment by pipetting up and down 2 times.
- 13. Gravity sediment the clusters for 2–5 minutes.
- While the colony fragments are sedimenting, aspirate the matrix solution from the freshly prepared rhLaminin-521 coated 60-mm dish and add 4 mL of Essential 8<sup>™</sup> Medium.
- Aspirate the supernatant and resuspend the sedimented PSC clusters by gently pipetting them up and down 2 times in 4 mL Essential 8<sup>™</sup> Medium, taking care not to break them down further.
- 16. Distribute 1 mL of the resuspended PSC clusters into the rhLaminin-521 pre-coated dish. Move the vessel in several quick back-and-forth and side-to-side motions to disperse the cells across its surface.
- 17. Incubate the cells in a 37°C, 5% CO<sub>2</sub> incubator and passage them when they are 60%–85% confluent to maintain optimal cell health.

**Note:** Cells cultured in Essential  $8^{M}$  Medium must be fed daily.

Table 1 rhLaminin-521 Coating Reagent volumes (per well or per dish)

Culture vessel (surface area)	Volume of diluted rhLaminin-521 solution
6-well (10 cm <sup>2</sup> )	2 mL
12-well (4 cm <sup>2</sup> )	0.8 mL
24-well (2 cm <sup>2</sup> )	0.4 mL
35-mm (10 cm <sup>2</sup> )	2 mL
60-mm (20 cm <sup>2</sup> )	4 mL
100-mm (60 cm <sup>2</sup> )	12 mL

**Table 2** Passaging and culture reagent volumes (per well or per dish)

Culture vessel (surface area)	DPBS for wash	1 mg/mL Collagenase IV	Essential 8™ Medium*	Essential 8™ Medium**
6-well (10 cm <sup>2</sup> )	2 mL	1 mL	1 mL	2 mL
12-well (4 cm <sup>2</sup> )	1 mL	0.4 mL	0.4 mL	8 mL
24-well (2 cm <sup>2</sup> )	0.5 mL	0.2 mL	0.2 mL	0.4 mL
35-mm (10 cm <sup>2</sup> )	2 mL	1 mL	1 mL	2 mL
60-mm (20 cm <sup>2</sup> )	4 mL	2 mL	2 mL	4 mL
100-mm (60 cm <sup>2</sup> )	12 mL	6 mL	6 mL	12 mL

\*For initial resuspension and for wash off. \*\*For final two resuspensions. **Note**: Split ratios may need to be optimized depending on the cell line and the percentage confluency of PSCs at the time of harvest.

#### Related products

Product	Cat. no.
DPBS, calcium, magnesium	14040
Essential 8 <sup>™</sup> Medium	A1517001
rhLaminin-521	A29248
TrypLE <sup>™</sup> Select Enzyme (1X), no phenol red	12563
DPBS, no calcium, no magnesium	17104
Collagenase, Type IV, Powder	A16517
DMEM/F12, GlutaMAX <sup>™</sup> Supplement	10565

## Explanation of symbols and warnings

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Caution, consult accompanying documents	Temperature Limitation	Keep away from light	Use By:	Consult instructions for use
LOT	REF		Read SDS	
Batch Code	Catalog	Manufacturer	Read Safety Data	

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