Binvitrogen Ilife technologies

ThermoScript[™] Reverse Transcriptase

Cat. No. 12236-014 Cat. No. 12236-022 Conc. 15 U/µl

Size: 375 Units Size: 1,500 Units Store at -20°C in a non-frost free freezer

Description

ThermoScript[™] Reverse Transcriptase (RT) is an avian reverse transcriptase that has been engineered for reduced RNase H activity and higher thermal stability (1). It produces higher yields of cDNA and more full-length cDNA transcripts than AMV RT. ThermoScript[™] RT can generate cDNA transcripts from 100 bp to >12 kb at temperatures ranging from 50°C to 65°C.

Component

Component	375-U Kit	1,500-U Kit
ThermoScript [™] RT (15 U/μl)	25 µl	100 µl
5X cDNA Synthesis Buffer	500 µl	$2 \times 500 \ \mu l$
0.1 M DTT	100 µl	100 µl

Unit Definition

One unit incorporates 1 nmol of dTTP into acid-precipitable material in 10 min at 37°C using poly(A)•oligo(dT)25 as template-primer .

Storage Buffer

200 mM KPO₄ (pH 7.1), 0.1 mM EDTA, 1 mM DTT, 0.05% (v/v) Triton[®] X-100, 50% (v/v) glycerol, stabilizers

5X cDNA Synthesis Buffer

250 mM Tris acetate (pH 8.4 at room temperature), 375 mM potassium acetate, 40 mM magnesium acetate

Storage and Handling Store ThermoScript^m RT at -20°C in a non-frost-free freezer. Stability may be extended by storing at -70°C. Store the 5X cDNA Synthesis Buffer and 0.1 M DTT at -20°C. Thaw the solutions at room temperature just prior to use and refreeze immediately.

Part no. 12236.pps

Rev. date: 26 Sep 2003

This product is distributed for laboratory research only. CAUTION: Not for diagnostic use. The safety and efficacy of this product in diagnostic or other clinical uses has not been established.

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First-Strand cDNA Synthesis Using ThermoScript [™] R	d cDNA Synthesis Using ThermoSc	ript [™] RT
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1. Add the following components to a nuclease-free microcentrifuge tube:		
Oligo (dT) ₂₀ (50 µM) or	1 µl	
200–500 ng (dT) ₁₂₋₁₈ or		
50–250 ng random primers <i>or</i>		
10–20 pmole gene-specific prime		
10 pg to 5 μg total RNA <i>or</i> 10 pg to 500 ng of mRNA	x µl	
10 mM dNTP Mix	2 µl	
Sterile, distilled water	to 12 µl	
 Incubate mixture at 65°C for 5 min and then place on ice (optional). Collect the contents of the tube by brief centrifugation and add: 		
5X cDNA Synthesis Buffer	4 µl	
0.1 M DTT	1 µl	
RNaseOUT TM (40 units / μ l) (optional)	* 1 μl	
Sterile, distilled water	1 µl	
ThermoScript [™] RT (15 U/µl)**	1 µl	
*RNaseOUT [™] Recombinant Ribonuclease Inhibitor (Cat. No. 10777-019) is required if using <50 ng starting RNA.		
**If less than 1 ng of RNA is used, reduce the amount of ThermoScript [™] RT in the		
	ase the amount of sterile, distilled water to	
1.5μ l/reaction (2).	whata tuba at 25% for 10 min	
3. If you are using random primers, ind		
 Mix contents of the tube gently and incubate at 50°C for 30-60 min. (If you are using oligo(dT)₂₀ or gene-specific primers, you can incubate at 50–65°C.) 		

Terminate the reaction by heating at 85°C for 5 min.

Note that amplification of PCR targets >1 kb may require the removal of RNA complementary to the cDNA. To remove RNA complementary to the cDNA, add 1 μ l (2 units) of *E. coli* RNase H and incubate at 37°C for 20 min.

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PCR

The following is intended as a guideline and starting point when using first-strand cDNA in PCR with *Taq* DNA polymerase. The optimal concentration of Mg⁺⁺ will vary depending on the template and primer pair.

Use only 10% of the first-strand reaction for PCR. Higher volumes may not increase amplification and may result in decreased amounts of PCR product.

1. Add the following to a PCR tube:

10X PCR Buffer [200 mM Tris-HCl (pH 8.4), 500 mM KCl]	5 µl
50 mM MgCl ₂	1.5 μl
10 mM dNTP Mix	1 µl
Forward primer (10 μM)	1 µl
Reverse primer (10 µM)	1 µl
Taq DNA polymerase (5 U/ μ l)	0.4 µl
cDNA from first-strand reaction	2 µl
autoclaved, distilled water	to 50 µl

2. Mix gently and layer with 1–2 drops (~50 µl) of silicone oil. (Note: Silicone oil is unnecessary in thermal cyclers equipped with a heated lid.)

3. Heat reaction to 94°C for 2 min to denature.

4. Perform 15 to 40 cycles of PCR. Use the recommended annealing and extension conditions for your *Taq* DNA polymerase.

Quality Control

This product has passed the following quality control assays: SDS-polyacrylamide gel analysis for purity; functional absence of endodeoxyribonuclease, 3' and 5' exodeoxyribonuclease, and ribonuclease activities; yield and length of cDNA product.

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References

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2. Huang, L., Lee, J., Sitaraman, K., Gallego, A., and Rashtchian, A. (2000)

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