

Technical Data Sheet

DimerX I: PE Soluble Dimeric Mouse H-2Kb:Ig Fusion Protein

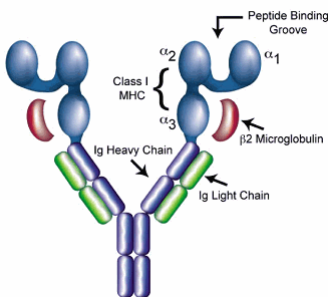
Product Information

Material Number:	552944
Size:	0.1 mg
Concentration:	0.2 mg/ml
Isotype:	Mouse IgG1, λ
Storage Buffer:	Aqueous buffered solution containing $\leq 0.09\%$ sodium azide.

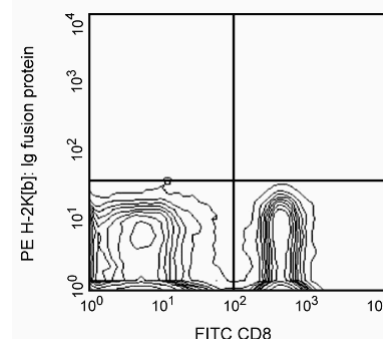
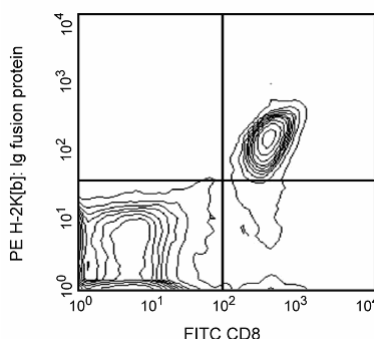
Description

The H-2K[b]:Ig fusion protein consists of three extracellular major histocompatibility complex (MHC) class I H-2Kb domains that are fused to the VH regions of mouse IgG1. In order for the MHC class I to be functional, i.e., capable of binding peptides, $\beta 2$ Microglobulin ($\beta 2$ M) must be present. For this reason, BD™ DimerX consists of recombinant H-2K[b]:Ig fusion protein, supplemented with recombinant $\beta 2$ M. Recombinant MHC molecules, like the BD DimerX fusion protein, are useful for studying T-cell function by immunofluorescent staining and flow cytometric analysis of antigen-specific T cells.

The MHC gene locus encodes a group of highly polymorphic, cell-surface proteins that play a broad role in the immune response to protein antigens. MHC molecules function by binding and presenting small antigenic protein fragments to antigen-specific receptors expressed by T cells (TCR). Human (human leukocyte antigen/HLA) and mouse (histocompatibility 2/H-2) MHC molecules are structurally and functionally related proteins that comprise two major classes. Class I MHC molecules consist of two separate polypeptide chains. The class I α chain is an MHC encoded, transmembrane polypeptide containing three extracellular domains: $\alpha 1$, $\alpha 2$, and $\alpha 3$. The second chain consists of a non-MHC encoded polypeptide called $\beta 2$ M. Since $\beta 2$ M does not contain a transmembrane domain, it associates with the α chain through noncovalent interaction. Functionally, class I MHC molecules can bind peptides derived from intracellular antigens (e.g., viral and some bacterial antigens) that are specifically recognized by CD8+ T cells. Class II MHC molecules consist of two different transmembrane proteins that can bind peptide fragments derived from extracellular proteins (e.g., bacteria and fungi) and are specifically recognized by CD4+ T cells. TCR recognize both processed peptides bound to MHC, as well as regions of the MHC molecule itself. CD4 and CD8 accessory molecules strengthen formation of the TCR-MHC complex through their interaction with nonpolymorphic regions of the MHC molecule.



Schematic representation of the MHC class I:Ig dimeric protein.



Antigen-specific T-cell identification of OT-1 TCR transgenic mouse splenocytes. Splenocytes from OT-1 TCR transgenic mice were incubated with Mouse BD Fc Block™ purified anti-mouse CD16/CD32 mAb 2.4G2 (Cat. No. 553141/553142, both panels) and stained with FITC-conjugated anti-mouse CD8a mAb 53-6.7 (Cat. No. 553030/553031, both panels) and PE-conjugated DimerX H-2Kb:Ig Fusion Protein (both panels) loaded overnight at 4°C with OVA peptide (left panel) or SIY peptide (right panel). Data shown is for viable cells only (10% probability plot). Cells were gated based on propidium iodide staining, as well as light-scatter characteristics. Data was acquired on a BD FACSCalibur™ flow cytometry system.

Preparation and Storage

Store undiluted at 4°C and protected from prolonged exposure to light. Do not freeze.

The H-2Kb:Ig Dimeric protein was expressed together with human $\beta 2$ M in the mouse plasmacytoma cell line, J558L (ATCC TIB-6). The H-2Kb and $\beta 2$ M polypeptide chains are associated noncovalently as a consequence of their coexpression within J558L cells. The H-2Kb:Ig fusion protein was purified from tissue culture supernatant by affinity chromatography. The protein was conjugated with R-PE under optimum conditions, and unconjugated protein and free PE were removed.

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Application Notes

Application

Flow cytometry	Routinely Tested
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Recommended Assay Procedure:

This H-2Kb:Ig fusion protein has been tested by immunofluorescent staining ($\leq 4 \mu\text{g}$ H-2Kb:Ig/million cells) and flow cytometric analysis of antigen-specific T cells to assure specificity and reactivity. It is necessary to load the H-2Kb portions of the dimeric protein with a relevant peptide of interest prior to immunofluorescent staining of T cells. H-2Kb:Ig complexes are effectively loaded by incubation with excess relevant (specific) or irrelevant (control) peptides (see Protocol 1). Peptide-loaded H-2Kb:Ig may be used for immunofluorescent staining (see Protocol 2). **Since applications vary, each investigator must determine dilutions appropriate for individual use.**

Protocol 1: Peptide Loading of H-2Kb:Ig Dimeric Protein

An alloreactive T-cell clone, 2C, is specific for an endogenous peptide, p2Ca. Several related peptides or peptide analogs have been identified. These differ in their MHC restriction and in their affinity for 2C TCR. For H-2Kb, SIY peptide can also be recognized by 2C TCR in the context of H-2Kb. SIY has a relatively high affinity for H-2Kb and is suggested as a positive control for staining of 2C cells in this assay. The 2C clone was originally derived by stimulating BALB/c spleen cells with irradiated P815 (H-2Ld) cells.

Several peptide-loading protocols have been described. The method used at BD Biosciences Pharmingen involves passive loading of excess peptide in solution with H-2Kb:Ig protein. We have found that passive loading works particularly well in the case of high-affinity peptides. For lower-affinity peptides, an increase in the molar ratio of peptide to H-2Kb:Ig may improve loading, as determined by flow cytometric analysis. It is suggested that for each peptide, parameters such as the dose of H-2Kb:Ig per million cells, molar ratio of peptide to H-2Kb:Ig, and peptide loading time be determined empirically by the investigator. While this DimerX product contains $\beta 2$ Microglobulin, for investigators requiring excess recombinant Human $\beta 2$ Microglobulin, we recommend BD Biosciences Cat. No. 551089.

Peptide preparation and loading:

1. The molecular weight (MW) of a peptide of interest will need to be determined. A peptide's MW can be estimated by multiplying its number (n) of amino acids (AA) by 130 daltons (d) per amino acid: **MW of peptide (d) = n (AA) x 130 (d/AA)**
2. A stock of peptide may be prepared at 20 mg/ml in DMSO. Dilute the peptide solution to 2 mg/ml in sterile DPBS, pH 7.2 for use in the H-2Kb:Ig loading protocol.
3. Mix H-2Kb:Ig protein with specific or control peptide at 40, 160, or 640 molar (M) excess.

The following calculation, using an 8 amino acid peptide (8mer) as an example, may be used:

Dp = Molecular Weight of peptide: eg, 8 amino acids x 130 = 1,040 daltons.

DKb = Molecular Weight of H-2Kb:Ig = 250,000 daltons.

R = desired excess molar ratio, eg, 160.

Mp = micrograms (μg) peptide of interest.

MKb = micrograms (μg) H-2Kb:Ig in the reaction. A typical amount of peptide-loaded H-2Kb:Ig to use for flow cytometry staining is 0.25 to 4 μg /million cells (test).

$$\text{Mp} = \frac{\text{MKb} \times \text{R} \times \text{Dp}}{\text{DKb}} = \frac{4 \mu\text{g} \times 160 \times 1,040 \text{ d}}{250,000 \text{ d}} = 2.66 \mu\text{g}$$

Therefore, one would add 2.66 μg of peptide and 4 μg of H-2Kb:Ig in solution for the optimal peptide loading of H-2Kb:Ig.

4. Mix peptide and H-2Kb:Ig together in PBS, pH 7.2, incubate at 37°C overnight. The peptide-loaded H-2Kb:Ig can be stored at 4°C for up to 1 week.

Protocol 2: Immunofluorescent Staining Protocol

1. Resuspend mouse cells in FACS staining buffer [e.g., DPBS, 1% FCS, 0.09% NaN₃ or BD Pharmingen™ Stain Buffer (FBS), Cat. No. 554656], containing the appropriate amount of Mouse BD Fc Block™ purified anti-mouse CD16/CD32 mAb 2.4G2 (Cat. No. 553141/553142), at a concentration of approximately 10e6 cells per 50 μl . Incubate 10 minutes at 4°C. Add $\sim 1 \times 10^6$ cells per staining tube (e.g., 12 x 75 mm tube, BD Falcon™ Cat. No. 352008).
2. Add 0.25 to 4 μg of peptide-loaded H-2Kb:Ig protein to cell suspension. Any cell-surface marker-specific antibodies can also be added to the cell suspension during this step. Incubate 60 minutes at 4°C.
3. Wash cells 2x with 2 ml FACS buffer, centrifuge for 5 minutes at 250 x g, and discard supernatant. Resuspend cell pellet in approximately 0.5 ml staining buffer in a tube appropriate for the flow cytometer.

Suggested Companion Products

Catalog Number	Name	Size	Clone
551089	Recombinant Human $\beta 2$ Microglobulin	0.1 mg	(none)
553030	FITC Rat Anti-Mouse CD8a	0.1 mg	53-6.7
553141	Purified Rat Anti-Mouse CD16/CD32 (Mouse BD Fc Block™)	0.1 mg	2.4G2
554656	Stain Buffer (FBS)	500 ml	(none)

Product Notices

1. Since applications vary, each investigator should titrate the reagent to obtain optimal results.
2. Please refer to www.bdbiosciences.com/pharming/en/protocols for technical protocols.
3. For fluorochrome spectra and suitable instrument settings, please refer to our Fluorochrome Web Page at www.bdbiosciences.com/colors.
4. Caution: Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.

References

- Cai Z, Brunmark AB, Luxembourg AT, et al. Probing the activation requirements for naive CD8+ T cells with Drosophila cell transfectants as antigen presenting cells. *Immunol Rev.* 1998; 165:249-265.(Biology)
- Cai Z, Kishimoto H, Brunmark A, Jackson MR, Peterson PA, Sprent J. Requirements for peptide-induced T cell receptor downregulation on naive CD8+ T cells. *J Exp Med.* 1997; 185(4):641-651.(Biology)
- Dal Porto J, Johansen TE, Catipovic B, et al. A soluble divalent class I major histocompatibility complex molecule inhibits alloreactive T cells at nanomolar concentrations. *Proc Natl Acad Sci U S A.* 1993; 90(14):6671-6675.(Biology)
- Schneck JP, Slansky JE, O'Herrin SM, Greten TF. Monitoring antigen-specific T cells using MHC-Ig dimers. In: Coligan J, Kruisbeek D, Margulies EM, Shevach EM, Strober W, ed. *Current Protocols in Immunology*. New York: John Wiley & Sons, Inc; 2000:17.2.1-17.2.17.(Biology)