Certificate of Analysis

pGL4.34[*luc2P*/SRF-RE/Hygro] Vector

Part No.	Size
E135A	20µg

> **Instructions for use** of this product can be found in the pGL4 Luciferase Reporter Vectors Technical Manual #TM259, available online at:
> **www.promega.com/tbs**

Description: The pGL4.34[*luc2P*/SRF-RE/Hygro] Vector[a–e] contains a Serum Response Factor Response Element (SRF-RE) that drives the transcription of the luciferase reporter gene *luc2P* in response to activation of Serum Response Factor through multiple signaling pathways including activation of RhoA GTPase. *luc2P* is a synthetically derived luciferase sequence with humanized codon optimization. The *luc2P* gene also contains hPEST, a protein destabilization sequence. The protein encoded by *luc2P* responds more quickly to induction than the protein encoded by the *luc2* gene. The vector backbone contains an ampicillin resistance gene to allow for selection in *E. coli* and the mammalian-selectable marker for hygromycin resistance.

Concentration: 1µg/µl.

GenBank® Accession Number: FJ773213.

Storage Buffer: The pGL4.34[*luc2P*/SRF-RE/Hygro] Vector is supplied in 10mM Tris-HCl, 1mM EDTA (pH 8.0).

Storage Conditions: See the Product Information Label for storage temperature recommendations and expiration date.

Usage Note: Mix well prior to use.

Quality Control Assays

Contaminant Assays

Contaminating Nucleic Acid Assay: RNA, single-stranded DNA and chromosomal DNA are not evident in a specified sample of this vector as determined by agarose gel electrophoresis.

Nuclease Assay: Following incubation of 1µg of this vector in Restriction Enzyme Buffer at 37°C for 16-24 hours, no evidence of nuclease activity is detected by agarose gel electrophoresis.

Physical Purity: $A_{260}/A_{280} \geq 1.80$, $A_{260}/A_{250} \geq 1.05$.

Functional Assays

Identity Assay: The vector has been sequenced completely and has 100% identity with the published sequence available at: **www.promega.com/vectors**

Restriction Digestion: The functional purity of this vector DNA is verified by complete digestion with several restriction enzymes at 37°C for 1 hour. Samples are examined by agarose gel electrophoresis, and cut and uncut vector DNA are compared with marker DNA.

Signed by: *J. Stevens*

J. Stevens, Quality Assurance

[a]READ THIS FIRST BEFORE OPENING PRODUCT
The sale of this product and its use by the purchaser are subject to the terms of a limited use label license, the full text of which is shipped with this product and also available at: **www.promega.com\LULL**. That text must be read by the purchaser prior to opening this product to determine whether the purchaser agrees that all use of the product shall be in accordance with the license terms. If the purchaser is not willing to accept the terms of the limited use label license, Promega is willing to accept the return of the unopened product and provide the purchaser with a full refund. However, if the product is opened for any reason, then the purchaser agrees to be bound by the terms of the limited use label license.
[b]Australian Pat. No. 2001 285278 and other patents pending.
[c]U.S. Pat. No. 5,670,356.
[d]Australian Pat. No. 2003272419 and other patents pending.
[e]The method of recombinant expression of *Coleoptera* luciferase is covered by U.S. Pat. Nos. 5,583,024, 5,674,713 and 5,700,673. A license (from Promega for research reagent products and from The Regents of the University of California for all other fields) is needed for any commercial sale of nucleic acid contained within or derived from this product.

PRODUCT USE LIMITATIONS, WARRANTY, DISCLAIMER

Features List and Map for the pGL4.34[*luc2P*/SRF-RE/Hygro] Vector

|	
SRF-RE	33–147
minimal promoter (minP)	180–210
luc2P reporter gene	243–2018
SV40 late poly(A) region	2058–2279
SV40 early enhancer/promoter	2327–2745
synthetic hygromycin (Hyg^r) coding region	2770–3807
synthetic poly(A) signal	3831–3879
reporter vector primer 4 (RVprimer4) binding region	3946–3965
ColE1-derived plasmid replication origin	4203
synthetic β-lactamase (Amp^r) coding region	4994–5854
synthetic poly(A) signal/transcriptional pause site	5959–6112
reporter vector primer 3 (RVprimer3) binding region	6061–6080
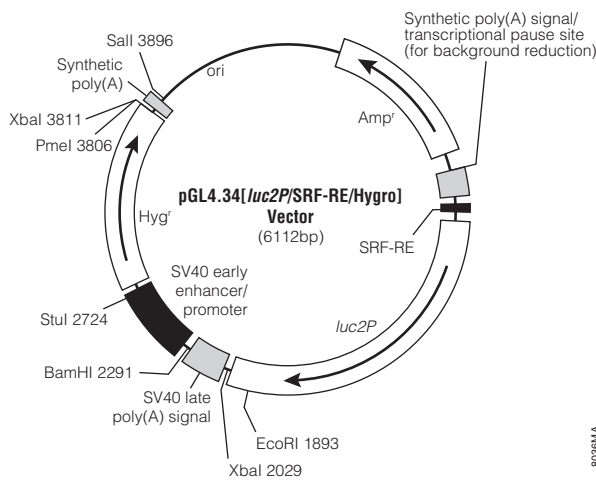
Figure 1. pGL4.34[*luc2P*/SRF-RE/Hygro] Vector map.

Sequence information, vector maps and restriction enzyme tables for the pGL4 Vectors are available online at: **www.promega.com/vectors**

Additional information is available in Technical Manual #TM259, available online at: **www.promega.com/tbs** or by request.

Sample Protocol to Determine Induction of Luciferase by FBS in HEK293 Cells Transfected with the pGL4.34[*luc2P*/SRF-RE/Hygro] Vector

Materials to Be Supplied by the User
- Dulbecco's PBS (DPBS)
- 0.05% (w/v) trypsin in DPBS
- DMEM
- DMEM supplemented with , 0.5%, 10% and 40% fetal bovine serum (DMEM/FBS)
- ONE-Glo™ Luciferase Assay System (Cat.# E6110)
- HEK293 cells
- transfection reagent

Day 1: Plate Cells

1. Grow HEK293 cells in DMEM/FBS to approximately 75% confluency.

2. Harvest cells via trypsinization: Remove the DMEM/FBS, wash the cells with DPBS, and add the trypsin/DPBS (1X volume). After 2 minutes, add a 4X volume of DMEM/FBS, collect the cell suspension and pellet the cells by centrifugation. Aspirate the supernatant and resuspend in DMEM/FBS. We have routinely used a concentration of 10,000–15,000 viable cells/100µl DMEM/FBS.

3. Dispense 100µl of the cell suspension into the wells of a 96-well plate. Plate enough wells to perform each test condition in triplicate.

4. Cover the plate and place it in a tissue culture incubator at 37°C overnight (or for 24 hours).

Day 2: Transfect Cells

1. Transfect the cells using a high-efficiency transfection reagent. Each well of 96-well plate to be transfected requires 0.1µg pGL4.34[*luc2P*/SRF-RE/Hygro] Vector plasmid DNA. Transfection conditions may require optimization.

2. Cover the plate and place it in a tissue culture incubator at 37°C.

3. After 4–6 hours, change the medium to DMEM/0.5%FBS (100µl per well) to start serum starvation.

Day 3: Induce Transfected Cells

1. Prepare 2X induction and 2X control solutions. Calculate the volume of 2X induction and 2X control solution by multiplying the number of wells needed for each solution by 50µl, and prepare 110% of this amount.
   - 2X induction solution: 40%FBS in DMEM
   - 2X control solution: DMEM

2. Remove 50µl media from wells that will be treated with either 2X induction solution or 2X control solution.

3. Add 50µl of 2X induction solution to the cells to be induced and 50µl of 2X control solution to the control noninduced cells.

4. Return the plate to the tissue culture incubator and induce for 6 hours.

5. Analyze luciferase activity using an appropriate luciferase detection assay. We have observed comparable results for fold induction of the vector using a variety of luciferase reagents, including: Bright-Glo™ Luciferase Assay System (Cat.# E2610, Technical Manual #TM052); ONE-Glo™ Luciferase Assay System (Cat.# E6110, Technical Manual #TM292); Dual-Luciferase® Reporter Assay System (Cat.# E1910, Technical Manual #TM040); and Dual-Glo® Luciferase Assay System (Cat.# E2920, Technical Manual #TM058).

6. Calculate the fold induction as follows:

$$\text{fold induction} = \frac{\text{average relative light units of induced cells}}{\text{average relative light units of control cells}}$$