

## DESCRIPTION

<b>Species Reactivity</b>	Human/Mouse/Rat
<b>Specificity</b>	Detects endogenous human, mouse, and rat full length and mitochondria-processed HTRA2/Omi.
<b>Source</b>	Polyclonal Rabbit IgG
<b>Purification</b>	Antigen Affinity-purified
<b>Immunogen</b>	<i>E. coli</i> -derived recombinant human HTRA2/Omi Ala134-Glu458 Accession # O43464
<b>Formulation</b>	Lyophilized from a 0.2 µm filtered solution in PBS with Trehalose. See Certificate of Analysis for details.

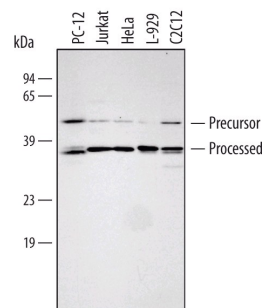
## APPLICATIONS

**Please Note:** Optimal dilutions should be determined by each laboratory for each application. [General Protocols](#) are available in the Technical Information section on our website.

	<b>Recommended Concentration</b>	<b>Sample</b>
<b>Western Blot</b>	0.25 µg/mL	See Below
<b>Immunocytochemistry</b>	5-15 µg/mL	See Below

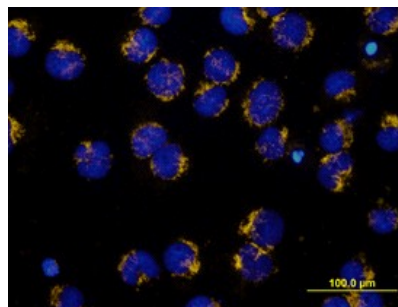
## DATA

### Western Blot



**Detection of Human/Mouse/Rat HTRA2/Omi by Western Blot.** Western blot shows lysates of PC-12 rat adrenal pheochromocytoma cell line, Jurkat human acute T cell leukemia cell line, HeLa human cervical epithelial carcinoma cell line, L-929 mouse fibroblast cell line, and C2C12 mouse myoblast cell line. PVDF membrane was probed with 0.25 µg/mL of Human/Mouse/Rat HTRA2/Omi Antigen Affinity-purified Polyclonal Antibody (Catalog # AF1458) followed by HRP-conjugated Anti-Rabbit IgG Secondary Antibody (Catalog # HAF008). Specific bands were detected for HTRA2/Omi at approximately 36 and 49 kDa (as indicated). This experiment was conducted under reducing conditions and using Immunoblot Buffer Group 2.

### Immunocytochemistry



**HTRA2/Omi in Jurkat Human Cell Line.** HTRA2/Omi was detected in immersion fixed Jurkat human acute T cell leukemia cell line stimulated with staurosporin using Human/Mouse/Rat HTRA2/Omi Antigen Affinity-purified Polyclonal Antibody (Catalog # AF1458) at 10 µg/mL for 3 hours at room temperature. Cells were stained using the NorthernLights™ 557-conjugated Anti-Rabbit IgG Secondary Antibody (yellow; Catalog # NL004) and counterstained with DAPI (blue). View our protocol for [Fluorescent ICC Staining of Cells on Coverslips](#).

## PREPARATION AND STORAGE

<b>Reconstitution</b>	Reconstitute at 0.2 mg/mL in sterile PBS.
<b>Shipping</b>	The product is shipped at ambient temperature. Upon receipt, store it immediately at the temperature recommended below.
<b>Stability &amp; Storage</b>	<b>Use a manual defrost freezer and avoid repeated freeze-thaw cycles.</b> <ul style="list-style-type: none"> <li>12 months from date of receipt, -20 to -70 °C as supplied.</li> <li>1 month from date of receipt, 2 to 8 °C, reconstituted.</li> <li>6 months from date of receipt, -20 to -70 °C, reconstituted.</li> </ul>

## BACKGROUND

HtrA2/Omi is the mammalian homologue of bacterial high temperature requirement protein (HtrA). HtrA2/Omi localizes to the mitochondria and is processed to expose an amino-terminal Reaper-like motif similar to SMAC/Diablo. HtrA2/Omi is released from the mitochondria in response to apoptotic insult and can interact with the BIR2 or BIR3 domains of XIAP to relieve caspase-IAP inhibition. This effect can be measured by reversing XIAP-BIR2 (R&D Systems, Catalog # 786-XB) inhibition of Caspase-7 (R&D Systems, Catalog # 823-C7) cleavage of a fluorogenic peptide (DEVD-AFC, MP Bio, Catalog # AFC-138). IC<sub>50</sub> values for this effect are typically between 0.2 and 1.5 µM. HtrA2/Omi is trimeric and functions as a serine protease. The serine protease activity may play a more central role in apoptosis than its IAP antagonizing function. A PDZ domain regulates the serine protease activity by blocking access to the active site. The specificity of the protease is yet to be defined and no endogenous substrates are known to date.

## References:

1. Suzuki, Y. *et al.* (2001) *Mol. Cell.* **8**:613.
2. van Loo, G. *et al.* (2002) *Cell Death & Diff.* **9**:20.
3. Hedge, R. *et al.* (2001) *J. Biol. Chem.* **277**:432.
4. Verhagen, A. *et al.* (2001) *J. Biol. Chem.* **277**:445.
5. Martins, L. *et al.* (2002) *J. Biol. Chem.* **277**:439.
6. Silke, J., and A. Verhagen (2002) *Cell Death & Diff.* **9**:362.
7. Savopoulos, J. *et al.* (2000) *Protein Expression & Purification* **19**:227.