

Platinum® Pfx DNA Polymerase

Cat. No. 11708-013 Size: 100 reactions 11708-021 250 reactions

11708-039 500 reactions

Conc: 2.5 U/µl Store at -20°C

Description

Platinum® *Pfx* DNA Polymerase is a proprietary enzyme preparation containing recombinant DNA polymerase from *Thermococcus* species strain KOD (1,2). Platinum® *Pfx* DNA Polymerase possesses proofreading 3′ to 5′ exonuclease activity and is a highly processive enzyme with fast chain extension capability (3).

Platinum® *Pfx* DNA Polymerase is provided in inactive form, due to specific binding of the Platinum® antibody. Polymerase activity is restored after a PCR denaturation step at 94°C, providing an automatic "hot start" for increased specificity, sensitivity, and yield (4). Platinum® *Pfx* DNA Polymerase is ideal for demanding PCR applications such as site-directed mutagenesis and PCR expression cloning.

For problematic and/or GC-rich templates, PCR $_{\rm x}$ Enhancer Solution is included with each kit (see the guidelines for use on page 2). The number of reactions per kit is based on a standard reaction size of 50 μ l.

Unit Definition

One unit of Platinum® *Pfx* incorporates 10 nmol of deoxyribonucleotide into acid-precipitable material in 30 minutes at 74°C.

Kit Components	100 Rxns	250 Rxns	500 Rxns
Platinum® <i>Pfx</i> DNA Polymerase	100 units	250 units	500 units
50 mM Magnesium Sulfate	1 ml	1 ml	1 ml
10X Pfx Amplification Buffer	1 ml	$2 \times 1 \text{ ml}$	$3 \times 1 \text{ ml}$
10X PCR _x Enhancer Solution	1 ml	$2 \times 1 \text{ ml}$	$3 \times 1 \text{ ml}$

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Important Recommendations and Guidelines

- Platinum[®] Pfx DNA Polymerase will not work in reactions that contain dUTP in either the dNTP mix or the primers.
- Use a 1X final concentration of 10X Pfx Amplification Buffer as a general starting point. A higher final buffer concentration (1.5X–2X) may enhance reaction yield and specificity.
- We recommend using 1 unit of Platinum[®] Pfx for most targets. For targets above 2 kb, up to 1.25 units may be used. Note that more enzyme may be inhibitory.
- The recommended starting concentration of MgSO₄ is 1 mM.
- Use an annealing temperature of 55–60°C as a general starting point.
- Only use PCR_x Enhancer Solution for problematic and/or GC-rich templates.

Guidelines for Using PCR_x Enhancer Solution

 PCR_x Enhancer Solution is included as an optional component for problematic and/or GC-rich templates. Use PCR_x Enhancer Solution in combination with 10X Pfx Amplification Buffer, *not* as a substitute.

PCR $_{\rm x}$ Enhancer Solution lowers the DNA melting temperature ($T_{\rm m}$), reducing the maximum primer annealing temperature approximately 2°C per 1X PCR $_{\rm x}$ Enhancer Solution concentration, while at the same time expanding the effective annealing temperature over a much broader range. To determine optimal concentrations and conditions, start with an annealing temperature of 55–60°C and varying the amount of 10X PCR $_{\rm x}$ Enhancer Solution. For targets with higher GC content (60–90%), test 10X PCR $_{\rm x}$ Enhancer Solution at final concentrations of 0.5X, 1X, 2X, and 3X.

Quality Control

The Certificate of Analysis (CofA) provides detailed quality control information for each product. CofAs are available on our website. Go to www.invitrogen.com/support and search for the CofA by product lot number, which is printed on the box.

PCR Protocol

The following protocol is specific for Platinum® *Pfx* DNA Polymerase. Follow these instructions carefully for optimal performance.

 Add the following components to an sterile microcentrifuge tube at room temperature or on ice. The amounts are for a single reaction. Prepare a master mix of common components for multiple reactions.

Component	Volume	Final Concentration
10X Pfx Amplification Buffer	5–10 µl *	1X-2X
10 mM dNTP mixture	1.5 µl	0.3 mM each
50 mM MgSO ₄	1 µl	1 mM
Primer mix (10 µM each)	1.5 µl	0.3 µM each
Template DNA (10 pg – 200 ng)	≥1 µl	As required
Platinum [®] <i>Pfx</i> DNA Polymerase	$0.4~\mu l$	1 unit
Autoclaved, distilled water	to 50 µl	

^{*}Use 5 µl (1X final concentration) of buffer as a general starting point.

- Mix tube contents. Cap the tube and centrifuge briefly to collect the contents.
- Denature the template for 2–5 minutes at 94°C. Note that a longer denaturation time (up to 5 minutes) may increase yield and specificity.
- 4. Perform 25–35 cycles of PCR amplification as follows:

Three-step cycling Denature: 94°C for 15 seconds Anneal: 55°C for 30 seconds Extend: 68°C for 1 minute per kb Two-step cycling Denature: 94°C for 15 seconds Extend: 68°C for 1 minute per kb

Note: Two-step cycling can be used for long primers with high T_m.

- 5. Maintain the reaction at 4°C after cycling. Samples can be stored at -20°C until use.
- 6. Analyze the products by agarose gel electrophoresis.

References

- Takagi et. al, (1997) Appli. Environ. Microbiol., 63, 4504-4510.
- 2. Nishioka et. al, (2001) J. Biotechnol., 88, 141-9.
- 3. Cline et. al, (1996) Nucleic Acid Res., 24, 3546.
- 4. Sharkey et. al, (1994) BioTechnology, 12, 506.

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