

AccuPrime[™] Pfx SuperMix

Cat. No. 12344-040 Size: 200 Reactions

Store at -20°C in a non-frost-free freezer

Description

AccuPrime[™] *Pfx* SuperMix provides qualified reagents for the high fidelity amplification of DNA templates by polymerase chain reaction (PCR). It includes recombinant DNA polymerase from *Thermococcus* species KOD, anti-KOD antibodies, thermostable AccuPrime[™] proteins, MgSO₄, dNTPs, and stabilizers in a convenient and highly optimized SuperMix formulation for ease of reaction setup (1). It is suitable for targets up to 15 kb in length.

AccuPrime[™] Pfx DNA polymerase possesses a proofreading 3′ to 5′ exonuclease activity that provides higher fidelity than Pfu DNA polymerase (2). This highly processive enzyme is provided in an antibody-bound form that is inactive at ambient temperatures. The enzyme regains activity after the initial denaturation step at 94°C in PCR cycling, providing an automatic "hot start" that increases specificity, sensitivity, and yield, while allowing room temperature assembly (3).

Thermostable AccuPrimeTM proteins enhance specific primer-template hybridization during every cycle of PCR (4). The high specificity, fidelity, and yield offered by AccuPrimeTM Pfx SuperMix make it ideal for demanding PCR applications such as site-directed mutagenesis and PCR expression cloning.

AccuPrime $^{\text{\tiny M}}$ Pfx SuperMix is supplied at 1.1X concentration to allow approximately 10% of the final reaction volume to be used for the addition of primer and template solutions. Reagents sufficient for 200 amplification reactions of 25 µl each are provided.

<u>Component</u> AccuPrime[™] Pfx SuperMix **200-Rxn kit** 4 × 1.125 ml

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Unit Definition

One unit of AccuPrimeTM Pfx DNA Polymerase incorporates 10 nmol of deoxyribonucleotide into acid-insoluble material in 30 min at 74°C.

AccuPrime[™] Pfx SuperMix Components

22 U/ml *Thermococcus* species KOD thermostable polymerase complexed with anti-KOD antibodies, 66 mM Tris-SO₄(pH 8.4), 30.8 mM (NH₄)₂SO₄, 11 mM KCl, 1.1 mM MgSO₄, 330 μM dNTPs, AccuPrime[™] proteins, stabilizers.

Quality Control

The Certificate of Analysis (CofA) provides detailed quality control information for each product. The CofA is available on our website at www.invitrogen.com/cofa, and is searchable by product lot number, which is printed on each box.

Recommendations and Guidelines:

- PCR is a powerful technique capable of amplifying trace amounts of DNA; take all appropriate precautions to avoid cross-contamination.
- For multiple reactions, you can prepare a master mix of AccuPrime[™] Pfx SuperMix and the component(s) common to all reactions.
- The optimal annealing temperature should be 5–10°C lower than the T_m of the primers used; if necessary, gradually increase the annealing temperature by 2–3°C for higher specificity.
- If the PCR efficiency is not optimal, repeat the reaction with different primer concentrations from 100 to 500 nM, in 100 nM increments.

PCR Protocol

- 1. Add the following components in any order to each reaction tube:
 - 22.5 µl AccuPrime[™] Pfx SuperMix
 - Forward and reverse primers (200 nM final concentration of each is recommended)*
 - Template DNA solution (10 pg-200 ng)*
 - *A standard 25- μ l PCR reaction includes a combined primer and template volume of 2.5 μ l; we have observed no decrease in product yield if the amount of primer and template solution is between 0.5 μ l and 7.5 μ l.
- Mix contents of the tubes and overlay with mineral or silicone oil, if necessary.
- 3. Cap the tubes and load in the thermal cycler.
- Use the following PCR program as a starting point for your template and primers:

95°C for 5 minutes

35 cycles of:

95°C for 15 seconds

55-65°C for 30 seconds

68°C for 1 minute per kb

5. Maintain reaction at 4°C after cycling. Samples can be stored at -20°C.

References

- Takagi, M., Nishioka, M., Kakihara, H., Kitabayashi, M., Inoue, H., Kawakami, B., Oka, M., and Imanaka, T. (1997) Appli. Environ. Microbiol., 63, 4504-4510.
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- 3. Sharkey, D.J., Scalice, E.R., Christy, K.G., Atwood, S.M., Daiss, J.L. (1994) *BioTechnology*, 12, 506.
- 4. Rapley, R. (1994) Mol. Biotechnol., 2, 295-298.

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