

#### Thermo Scientific Thermo-Start Taq DNA Polymerase with High Performance Buffer

Thermo-Start<sup>TM</sup> Taq DNA Polymerase is a chemically modified **Description:** 

version of ThermoPrime Tag DNA Polymerase. It is completely inactive at room temperature, preventing the formation and subsequent amplification of non-specific products. The enzyme requires an

activation step at 95°C for 15 minutes.

**Enzyme Source:** Thermus aquaticus

**Concentration:** 5 units/µl

One unit of enzyme is defined as the amount that will incorporate **Unit Definition:** 

10nmoles of dNTPs into acid insoluble material in 30 minutes at 74°C

under the analysis conditions below.

Thermo-Start  $^{TM}$  has 5' to 3' polymerization and exonuclease activity but lacks 3' to 5' exonuclease activity (proofreading). Associated

**Activities:** 

Store Thermo-Start<sup>TM</sup> Taq DNA Polymerase at -20°C, in a constant Storage

temperature freezer for up to 12 months. Shipped on ice within the UK **Conditions:** 

and on dry ice for international and within the US.

#### Kit Contents

Vial	Pack Size (cap color)		
	A	В	C
Thermo-Start Taq DNA Pol.	1 x 50μ1	10 x 50μl	100 x 50μl
	(clear)	(clear)	(clear)
Thermo-Start High Performance	1 x 1.25ml	10 x 1.25ml	100 x 1.25ml
PCR Buffer	(red)	(red)	(red)
MgCl <sub>2</sub>	1 x 1.5ml	10 x 1.5ml	100 x 1.5ml
	(clear)	(clear)	(clear)



# Example of Protocol:

Mix and spin down the solutions prior to use

	Volume	Final Concentration 1X	
Thermo-Start Taq	0.125µl	0.625 U	
DNA Pol. (5U/μl)	0.125μ1		
10X Thermo-Start HP Buffer	2.5µ1	1X	
dNTP Mix (20mM)	1µl	0.5mM of each nucleotide	
MgCl <sub>2</sub> (25mM)	1.5µ1*	1.5mM*	
Primer forward (10µM each)	1.25µ1*	0.5μM*	
Primer reverse (10µM each)	1.25µl*	0.5μM*	
Water (PCR Grade)	variable		
DNA Template	0.5 - 10μ1	0.5 - 125ng	
Total Volume	25 µl		

<sup>\*</sup>Scale up or down the volume and concentration as appropriate

Note: These recommendations are intended as basic guidelines. Magnesium chloride concentration and amount of enzyme should be optimized according to template and primer combination.

# Example of Program:

	Temp.	Time	Number of cycle
Initial Denaturation	95°C	15 min	1 cycle
Denaturation	95°C	20 sec	30 to 40
Annealing	50-65°C	30 sec	cycles
Extension**	72°C	60 sec	Cycles
Final Extension	72°C	5 min	1 cycle

<sup>\*\*</sup>Increase length of time in proportion to size of amplicon, Taq DNA Polymerase extends at approximately 1000 bp/min.

### **Incremental Activation:**

For extra stringency, the enzyme can be activated gradually during the PCR in a series of steps. The initial activation step is replaced by longer (2 minutes) denaturation steps for the first 7–8 cycles of the reaction.



Analysis Conditions: 25mM TAPS, pH 9.3 (at 25°C)

[tris-(hydroxymethyl)-methyl-amino-propane sulfonic acid, sodium salt]

 $\begin{array}{ccc} 50 mM & KCl \\ 2 mM & MgCl_2 \end{array}$ 

1mM β-mercaptoethanol

 $\begin{array}{lll} 200\mu M & \text{of each: dATP, dGTP, dTTP} \\ 100\mu M & [\alpha^{32}P]\text{-dCTP } (0.05 \text{ to } 0.1 \text{ Ci/mmol}) \\ 1.25\mu g/\mu l & \text{activated salmon sperm DNA} \end{array}$ 

Water added to a total volume of  $50\mu l$ . Incubated at  $74^{\circ}C$  for 10 minutes. The enzyme is first treated with a 15 minute activation step at  $95^{\circ}C$ . The amount of incorporated dNTPs is determined by trichloroacetic acid precipitation.

Storage Buffer: 100mM KCl

20mM Tris-HCl, pH 9.2 (at 25°C)

0.1mM EDTA (ethylenediaminetetraacetic acid)

1mM DTT (dithiothreitol)

0.5% Tween® 20 0.5% Nonidet® P40 50% (v/v) Glycerol

#### Ordering Information:

AB-1057/A	Thermo-Start <i>Taq</i> DNA Polymerase with High Performance Buffer	250 units
AB-1057/B	Thermo-Start <i>Taq</i> DNA Polymerase with High Performance Buffer	10 x 250 units
AB-1057/C	Thermo-Start <i>Taq</i> DNA Polymerase with High Performance Buffer	100 x 250 units

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