MfeI







R0589S



Recognition Site:

5′... C^TA A T T G ... 3′ 3′... G T T A A_c C ... 5′

Source: An *E. coli* strain that carries the cloned Mfel gene from *Mycoplasma fermentas* (N.F. Halden)

Also Available In High-Fidelity (HF™) Format Supplied in: 50 mM NaCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM DTT, 200 μ g/ml BSA and 50% glycerol.

Reagents Supplied with Enzyme: 10X NFBuffer 4.

Reaction Conditions: 1X NEBuffer 4. Incubate at 37°C.

1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM DTT pH 7.9 at 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 μg of λ DNA in 1 hour at 37°C in a total reaction volume of 50 μl .

Diluent Compatibility: Diluent Buffer A 50 mM KCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM DTT, 200 μg/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

Quality Control Assays

Ligation: After 20-fold overdigestion with Mfel, > 95% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1–2 μ M) at 16°C. Of these ligated fragments, > 95% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 35 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

Exonuclease Activity: Incubation of 100 units of enzyme with 1 μ g sonicated 3 H DNA (10 5 cpm/ μ g) for 4 hours at 37 $^\circ$ C in 50 μ l reaction buffer released < 0.3% radioactivity.

Endonuclease Activity: Incubation of 70 units of enzyme with 1 μ g pUC19 plasmid DNA for 4 hours at 37°C in 50 μ l reaction buffer resulted in < 5% conversion to RF II.

Blue/White Screening Assay: This enzyme has been tested to determine the integrity of the DNA ends produced after digestion with an excess of enzyme. An appropriate vector is digested at a unique site within $lacZ^{x}$ gene with a 10-fold excess of enzyme, ligated, transformed and plated on XGal/IPTG/Amp plates. Successful expression of β -galactosidase is a function of how intact its gene remains after cloning, an intact gene gives rise to a blue colony, an interrupted gene (i.e. degraded DNA end) gives rise to a white colony. Enzymes must produce fewer than 3% white colonies to be Blue/White Certified.

Enzyme Properties

Activity in NEBuffers:

NEBuffer 1 75% NEBuffer 2 50% NEBuffer 3 10% NEBuffer 4 **100%**

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

(see other side)

CERTIFICATE OF ANALYSIS

MfeI



1-800-632-7799 info@neb.com www.neb.com



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Reaction Conditions: 1X NEBuffer 4. Incubate at 37°C.

1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM DTT pH 7.9 at 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 μ g of λ DNA in 1 hour at 37°C in a total reaction volume of 50 μ l.

Diluent Compatibility: Diluent Buffer A 50 mM KCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM DTT, 200 μ g/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

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Activity in NEBuffers: NEBuffer 1 75%

NEBuffer 2 50% NEBuffer 3 10% NEBuffer 4 **100%**

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Survival in a Reaction: A minimum of 0.25 unit is required to digest 1 μ g of substrate DNA in 16 hours.

Heat Inactivation: 65°C for 20 minutes.

Notes: MunI is an isoschizomer of Mfel.

Not sensitive to *dam*, *dcm* or mammalian CpG methylation.

Conditions of low ionic strength, high enzyme concentration, glycerol concentration > 5% or pH > 8.0 may result in star activity.

Companion Products:

Mfel-HF™

#R3589S 500 units #R3589L 2,500 units

Mfel-HF™ RE-Mix™

#R5589S 25 reactions

= Time-Saver™ Qualified (See www.neb.com for details).

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