

# KasI



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R0544S 040121113111

## R0544S



250 units 4,000 U/ml Lot: 0401211

RECOMBINANT Store at -20°C Exp: 11/13

### Recognition Site:

5'...GGCGCC...3'  
3'...CCGCGG...5'

**Source:** An *E. coli* strain that carries the cloned KasI gene from *Kluyvera ascorbata* (C. Polisson)

New Reaction Buffer

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**Source:** An *E. coli* strain that carries the cloned KasI gene from *Kluyvera ascorbata* (C. Polisson)

New Reaction Buffer

Supplied in: 500 mM KCl, 20 mM Tris-HCl (pH 7.0), 0.1 mM EDTA, 0.1% Triton X-100, 1 mM MgCl<sub>2</sub>, 200 µg/ml BSA and 50% glycerol.

**Note:** -80°C is recommended for storage longer than 6 months.

**Reagents Supplied with Enzyme:**  
10X NEBuffer 4, 100X BSA.

**Reaction Conditions:** 1X NEBuffer 4, supplemented with 100 µg/ml BSA. Incubate at 37°C.

**1X NEBuffer 4:**  
50 mM potassium acetate  
20 mM Tris-acetate  
10 mM magnesium acetate  
1 mM DTT  
pH 7.9 @ 25°C

**Unit Definition:** One unit is defined as the amount of enzyme required to digest 1 µg of pBR322 DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

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**Diluent Compatibility:** Diluent Buffer B 300 mM NaCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM DTT, 500 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

### Quality Control Assays

**Ligation:** After 20-fold overdigestion with KasI, > 95% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1-2 µM) at 16°C. Of these ligated fragments, > 95% can be recut.

**16-Hour Incubation:** A 50 µl reaction containing 1 µg of DNA and 4 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

**Exonuclease Activity:** Incubation of 10 units of enzyme with 1 µg sonicated <sup>3</sup>H DNA (10<sup>5</sup> cpm/µg) for 4 hours at 37°C in 50 µl reaction buffer released 0.8% radioactivity.

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**Blue/White Screening Assay:** This enzyme has been tested to determine the integrity of the DNA ends produced after digestion with an excess of enzyme. An appropriate vector is digested at a unique site within *lacZ*<sup>α</sup> gene with a 10-fold excess of enzyme, ligated, transformed and plated on XGal/IPTG/Amp plates. Successful expression of β-galactosidase is a function of how intact its gene remains after cloning, an intact gene gives rise to a blue colony, an interrupted gene (i.e. degraded DNA end) gives rise to a white colony. Enzymes must produce fewer than 3% white colonies to be Blue/White Certified.

### Enzyme Properties

#### Activity in NEBuffers:

NEBuffer 1	25%
NEBuffer 2	100%
NEBuffer 3	0%
NEBuffer 4	100%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

(See other side)

CERTIFICATE OF ANALYSIS

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**Survival in a Reaction:** Not recommended for digest over 1 hour.

**Heat Inactivation:** 20 units of enzyme were inactivated by incubation at 65°C for 20 minutes.

**Notes:** KasI is an isoschizomer of NarI and SfoI.

KasI produces a 4-base 5' extension whereas NarI produces a 2-base 5' extension. KasI demonstrates marked site preference and is 25-fold more active on  $\lambda$  DNA than on pBR322 DNA.

Cleavage of mammalian genomic DNA is blocked by CpG methylation.

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