













1.000 units

8.000 U/ml

Lot: 0291205

RECOMBINANT Store at -20°C Exp: 5/14

# **Recognition Site:**

5...CCANNNNNTGG...3 3′... G G T N,N N N N A C C ... 5′

**Source:** An *E. coli* strain that carries the cloned PfIMI gene from *Pseudomonas fluorescens* (R. Morgan)

Supplied in: 50 mM KCl, 10 mM Tris-HCl (pH 7.5), 0.1 mM EDTA, 1 mM DTT, 200 µg/ml BSA and 50% glycerol.

Reagents Supplied with Enzyme: 10X NEBuffer 3, 100X BSA.

Reaction Conditions: 1X NEBuffer 3, supplemented with 100 ug/ml BSA. Incubate at 37°C.

1X NEBuffer 3:

100 mM NaCl 50 mM Tris-HCI 10 mM MgCl<sub>a</sub> 1 mM DTT pH 7.9 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1  $\mu$ g of  $\lambda$  DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

**Diluent Compatibility:** Diluent Buffer A 50 mM KCl. 10 mM Tris-HCl. 0.1 mM EDTA. 1 mM DTT, 200 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

### **Quality Control Assays**

**Ligation:** After 10-fold overdigestion with PfIMI, approximately 75% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1–2  $\mu$ M) at 16°C. Of these

ligated fragments, approximately 75% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 ug of DNA and 40 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

**Exonuclease Activity:** Incubation of 100 units of enzyme with 1 μg sonicated 3H DNA (105 cpm/μg) for 4 hours at 37°C in 50 µl reaction buffer released < 0.1% radioactivity.

Endonuclease Activity: Incubation of 100 units of enzyme with 1 µg pNEB193 plasmid DNA for 4 hours at 37°C in 50 µl reaction buffer resulted in < 10% conversion to RF II.

## **Enzyme Properties** Activity in NEBuffers:

NEBuffer 1 0% NEBuffer 2 100% NEBuffer 3 100% NEBuffer 4 50%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

Survival in a Reaction: A minimum of 0.5 unit is required to digest 1 µg of substrate DNA in 16 hours.

Heat Inactivation: 65°C for 20 minutes.

**Notes:** Particular PfIMI sites in  $\lambda$  DNA are cleaved at significantly lower rates than those found in other substrates.

Blocked by overlapping dcm methylation.

Conditions of low ionic strength, high enzyme concentration, glycerol concentration > 5% or pH > 8.0 may result in star activity.

### **Companion Products:**

dam-/dcm- Competent E. coli

#C2925H 20 transformation reactions #C29251 24 transformation reactions

= Time-Saver™ Qualified (See www.neb.com for details).

CERTIFICATE OF ANALYSIS

Survival in a Reaction: A minimum of 0.5 unit is required to digest 1 µg of substrate DNA in 16 hours.

Heat Inactivation: 65°C for 20 minutes.

**Notes:** Particular PfIMI sites in  $\lambda$  DNA are cleaved at significantly lower rates than those found in other substrates.

Blocked by overlapping dcm methylation.

Conditions of low ionic strength, high enzyme concentration, glycerol concentration > 5% or pH > 8.0 may result in star activity.

### **Companion Products:**

dam-/dcm- Competent E. coli

#C2925H 20 transformation reactions #C2925 24 transformation reactions

= Time-Saver™ Qualified (See www.neb.com for details).

**PfIMI** 



1-800-632-7799 info@neb.com www.neb.com

**R0509S** 







8,000 U/ml Lot: 0291205

RECOMBINANT Store at -20°C Exp: 5/14

# **Recognition Site:**

1,000 units

5'... C C A N N N N T G G ... 3' 3′...GGTNNNNNACC...5′

**Source:** An *E. coli* strain that carries the cloned PfIMI gene from Pseudomonas fluorescens (R. Morgan)

Supplied in: 50 mM KCl, 10 mM Tris-HCl (pH 7.5), 0.1 mM EDTA, 1 mM DTT, 200 µg/ml BSA and 50% alvcerol.

## Reagents Supplied with Enzyme: 10X NEBuffer 3, 100X BSA.

Reaction Conditions: 1X NEBuffer 3, supplemented with 100 μg/ml BSA. Incubate at 37°C.

#### 1X NEBuffer 3:

100 mM NaCl 50 mM Tris-HCI 10 mM MgCl<sub>o</sub> 1 mM DTT pH 7.9 @ 25°C

**Unit Definition:** One unit is defined as the amount of enzyme required to digest 1  $\mu$ g of  $\lambda$  DNA in 1 hour at 37°C in a total reaction volume of 50 µl.

**Diluent Compatibility:** Diluent Buffer A 50 mM KCl. 10 mM Tris-HCl. 0.1 mM EDTA. 1 mM DTT, 200 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C)

## **Quality Control Assays**

**Ligation:** After 10-fold overdigestion with PfIMI, approximately 75% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1-2 µM) at 16°C. Of these

ligated fragments, approximately 75% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 ug of DNA and 40 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

Exonuclease Activity: Incubation of 100 units of enzyme with 1 µg sonicated 3H DNA (105 cpm/ μg) for 4 hours at 37°C in 50 μl reaction buffer released < 0.1% radioactivity.

Endonuclease Activity: Incubation of 100 units of enzyme with 1 µg pNEB193 plasmid DNA for 4 hours at 37°C in 50 µl reaction buffer resulted in < 10% conversion to RF II.

## **Enzyme Properties Activity in NEBuffers:**

NEBuffer 1 0% NEBuffer 2 100% NEBuffer 3 **100**% NEBuffer 4 50%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.