# Tth 1111



info@neb.com

www.neb.com



**R0185S** 



4.000 U/ml 400 units

Lot: 0341207

RECOMBINANT Store at -20°C Exp: 7/14

# **Recognition Site:**

5′... GACN NNGTC...3′ 3'... C T G N N N C A G ... 5'

Source: An E. coli strain that carries the cloned Tth1111 gene from *Thermus thermophilus* 111 (T. Oshima)

Supplied in: 500 mM NaCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol, 200 ug/ml BSA and 50% glycerol.

# Reagents Supplied with Enzyme: 10X NEBuffer 4.

Reaction Conditions: 1X NEBuffer 4.

Incubate at 65°C.

### 1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM dithiothreitol pH 7.9 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of pBC4 DNA in 1 hour at 65°C in a total reaction volume of 50 µl.

Diluent Compatibility: Diluent Buffer B 300 mM NaCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM dithiothreitol, 500 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C).

# **Quality Control Assays**

Ligation: After 4-fold overdigestion with Tth1111, approximately 25% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1-2 µM) at 16°C. Of these ligated fragments, > 95% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 4 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

**Exonuclease Activity:** Incubation of 100 units of enzyme with 1 ug sonicated [3H] DNA (105 cpm/ug) for 4 hours at 65°C in 50 ul reaction buffer released < 0.1% radioactivity.

Endonuclease Activity: Incubation of 2 units of enzyme with 1 ug pUC19 RF I DNA for 4 hours at 65°C in 50 ul reaction buffer resulted in < 20% conversion to RF II.

# **Enzyme Properties**

# **Activity in NEBuffers:**

NEBuffer 1 50% NEBuffer 2 25% NEBuffer 3 25% NEBuffer 4 100%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

Survival in a Reaction: A minimum of 0.25 unit is required to digest 1 µg of substrate DNA in 16 hours.

**Heat Inactivation:** No

Plasmid Cleavage: Number of units required to cleave 1 µg of supercoiled plasmid DNA in one hour: pBR322 = 2 units.

**Notes:** Tth1111 produces DNA fragments that have a single-base 5' extension which are more difficult to ligate than blunt-ended fragments.

Not sensitive to dam, dcm or mammalian CpG methylation.

Incubation at 37°C results in 10% activity.

Conditions of low ionic strength, high enzyme concentration, glycerol concentration > 5% or pH > 8.0 may result in star activity. PfIFI, an isoschizomer of Tth1111, does not exhibit star

The activity in NEBuffer 1 is sensitive to pH. Slightly acidic pH conditions can cause a dramatic decrease in activity.

= Time-Saver™ Qualified (See www.neb.com for details).

CERTIFICATE OF ANALYSIS

# Tth 1111



1-800-632-7799 info@neb.com www.neb.com

R0185S

RR C NEB 4 65° W/6

400 units

4.000 U/ml

Lot: 0341207

RECOMBINANT Store at -20°C Exp: 7/14

# **Recognition Site:**

5′... GACN NNGTC...3′ 3'... C T G N N N C A G ... 5'

**Source:** An *E. coli* strain that carries the cloned Tth1111 gene from Thermus thermophilus 111 (T. Oshima)

Supplied in: 500 mM NaCl, 10 mM Tris-HCl (pH 7.4), 0.1 mM EDTA, 1 mM dithiothreitol, 200 µg/ml BSA and 50% glycerol.

## Reagents Supplied with Enzyme: 10X NFBuffer 4

Reaction Conditions: 1X NEBuffer 4. Incubate at 65°C.

### 1X NEBuffer 4:

50 mM potassium acetate 20 mM Tris-acetate 10 mM magnesium acetate 1 mM dithiothreitol pH 7.9 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme required to digest 1 µg of pBC4 DNA in 1 hour at 65°C in a total reaction volume of 50 ul.

Diluent Compatibility: Diluent Buffer B 300 mM NaCl, 10 mM Tris-HCl, 0.1 mM EDTA, 1 mM dithiothreitol, 500 µg/ml BSA and 50% glycerol (pH 7.4 @ 25°C).

# **Quality Control Assays**

Ligation: After 4-fold overdigestion with Tth1111. approximately 25% of the DNA fragments can be ligated with T4 DNA Ligase (at a 5' termini concentration of 1-2 uM) at 16°C. Of these ligated fragments. > 95% can be recut.

16-Hour Incubation: A 50 µl reaction containing 1 µg of DNA and 4 units of enzyme incubated for 16 hours resulted in the same pattern of DNA bands as a reaction incubated for 1 hour with 1 unit of enzyme.

Exonuclease Activity: Incubation of 100 units of enzyme with 1 µg sonicated [3H] DNA (105 cpm/µg) for 4 hours at 65°C in 50 µl reaction buffer released < 0.1% radioactivity.

Endonuclease Activity: Incubation of 2 units of enzyme with 1 µg pUC19 RF I DNA for 4 hours at 65°C in 50 ul reaction buffer resulted in < 20% conversion to RF II.

# **Enzyme Properties**

# **Activity in NEBuffers:**

NEBuffer 1 50% NEBuffer 2 25% NEBuffer 3 25% NEBuffer 4 100%

When using a buffer other than the optimal (supplied) NEBuffer, it may be necessary to add more enzyme to achieve complete digestion.

Survival in a Reaction: A minimum of 0.25 unit is required to digest 1 µg of substrate DNA in 16 hours.

### **Heat Inactivation:** No

Plasmid Cleavage: Number of units required to cleave 1 µg of supercoiled plasmid DNA in one hour: pBR322 = 2 units.

**Notes:** Tth1111 produces DNA fragments that have a single-base 5' extension which are more difficult to ligate than blunt-ended fragments.

Not sensitive to dam, dcm or mammalian CpG methylation.

Incubation at 37°C results in 10% activity.

Conditions of low ionic strength, high enzyme concentration, glycerol concentration > 5% or pH > 8.0 may result in star activity. PfIFI, an isoschizomer of Tth1111. does not exhibit star activity.

The activity in NEBuffer 1 is sensitive to pH. Slightly acidic pH conditions can cause a dramatic decrease in activity.

= Time-Saver™ Qualified (See www.neb.com for details).