

**Crimson Taq
DNA Polymerase with
(Mg-free) Buffer**



M0325S



200 units 5,000 U/ml Lot: 0141203
RECOMBINANT Store at -20°C Exp: 3/14

Description: *Taq* DNA Polymerase is a thermostable DNA polymerase that possesses a 5' → 3' polymerase activity (1,2,3) and a 5' flap endonuclease activity (4,5).

Crimson *Taq* DNA Polymerase offers superior performance in its newly formulated PCR buffer. Crimson *Taq* (Mg-free) Reaction Buffer contains a density reagent, which allows direct loading of PCR products onto a gel. In addition, Crimson *Taq* (Mg-free) Reaction Buffer has trace amounts of a red dye, which serves as an indicator for homogenous reaction setup, a color aid in gel loading and a tracking dye which migrates at about 10 bp on a 1% TBE gel.

Source: An *E. coli* strain that carries the *Taq* DNA Polymerase gene from *Thermus aquaticus* YT-1.

Applications:

- Primer Extension
- Routine PCR
- Colony PCR
- DHPLC
- Microarray Analysis
- High-throughput PCR

Supplied in: 100 mM KCl, 10 mM Tris-HCl (pH 7.4 @ 25°C), 0.1 mM EDTA, 1 mM dithiothreitol, 0.5% Tween 20, 0.5% NP-40 and 50% glycerol

Reagents Supplied with Enzyme:

5X Crimson *Taq* (Mg-free) Reaction Buffer
25 mM MgCl₂

Reaction Conditions: 1X Crimson *Taq* (Mg-free) Reaction Buffer, DNA template, primers, 200 μM dNTPs (not included), 1.5 mM MgCl₂ and 1.25–2.5 units of Crimson *Taq* DNA Polymerase in a total reaction volume of 50 μl.

1X Crimson Taq (Mg-free) Reaction Buffer:

12.5 mM Tricine
42.5 mM KCl
6% Dextran
Acid Red
pH 8.5 @ 25°C

Unit Definition: One unit is defined as the amount of enzyme that will incorporate 15 nmol of dNTP into acid insoluble material in 30 minutes at 75°C.

Unit Assay Conditions: 1X ThermoPol Reaction Buffer, 200 μM dNTPs including [³H]-dTTP and 200 μg/ml activated Calf Thymus DNA.

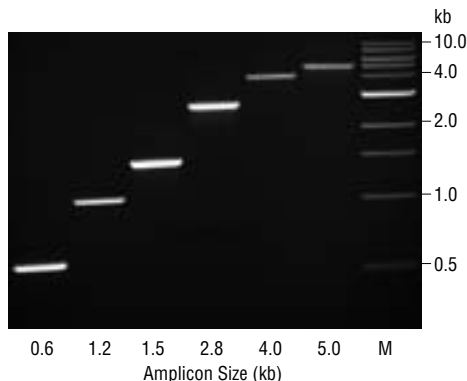
Heat Inactivation: No

Quality Control Assays

5 kb Lambda PCR: 25 cycles of PCR amplification of 5 ng Lambda DNA with 1.25 units of Crimson *Taq* DNA Polymerase in the presence of 200 μM dNTPs, 0.2 μM primers and 1.5 mM MgCl₂ in Crimson *Taq* (Mg-free) Reaction Buffer results in the expected 5 kb product.

3' → 5' Exonuclease Activity: Incubation of a 20 μl reaction in ThermoPol Reaction Buffer containing a minimum of 20 units of Crimson *Taq* DNA Polymerase with 10 nM fluorescent internally labeled oligonucleotide for 30 minutes at either 37°C or 75°C yields no detectable 3' → 5' degradation as determined by capillary electrophoresis.

Endonuclease Activity: Incubation of a 50 μl reaction in ThermoPol Reaction Buffer containing a minimum of 20 units of Crimson *Taq* DNA Polymerase with 1 μg of supercoiled φX174 DNA for 4 hours at 75°C results in < 10% conversion to the nicked form as determined by agarose gel electrophoresis.



Amplification of specific sequences from human genomic DNA using Crimson *Taq* DNA Polymerase. Amplicon sizes are indicated below gel. Marker M is NEB 1 kb DNA Ladder (NEB #N3232).

PCR

The Polymerase Chain Reaction (PCR) is a powerful and sensitive technique for DNA amplification (6). *Taq* DNA Polymerase is an enzyme widely used in PCR (7). The following guidelines are provided to ensure successful PCR using New England Biolabs' Crimson *Taq* DNA Polymerase. These guidelines cover routine PCR reactions. Amplification of templates with high GC content, high secondary structure, low template concentrations, or amplicons greater than 5 kb may require further optimization.

Reaction setup:

We recommend assembling all reaction components on ice and quickly transferring the reactions to a thermocycler preheated to the denaturation temperature (95°C).

COMPONENT	25 μl REACTION	50 μl REACTION	FINAL CONCENTRATION
5X Crimson <i>Taq</i> (Mg-free) Reaction Buffer	5 μl	10 μl	1X
25 mM MgCl ₂	1.5 μl	3 μl	1.5 mM
10 mM dNTP	0.5 μl	1 μl	200 μM
10 μM Forward Primer	0.5 μl	1 μl	0.2 μM (0.05–1 μM)
10 μM Reverse Primer	0.5 μl	1 μl	0.2 μM (0.05–1 μM)
Crimson <i>Taq</i> DNA Polymerase	0.125 μl	0.25 μl	1.25 units/ 50 μl PCR
Template DNA	variable	variable	<1,000 ng
Nuclease-Free Water	to 25 μl	to 50 μl	

Notes: Gently mix the reaction. Collect all liquid to the bottom of the tube by a quick spin if necessary. Overlay the sample with mineral oil if using a PCR machine without a heated lid.

Transfer PCR tubes from ice to a PCR machine with the block preheated to 95°C and begin thermocycling:

Thermocycling Conditions for a Routine PCR:

STEP	TEMP	TIME
Initial Denaturation	95°C	30 seconds
30 Cycles	95°C	15–30 seconds
	45–68°C	15–60 seconds
	68°C	1 minute/kb
Final Extension	68°C	5 minutes
Hold	4–10°C	

General Guidelines:

1. **Template:**
Use of high quality, purified DNA templates greatly enhances the success of PCR reactions. Recommended amounts of DNA template for a 50 μl reaction are as follows:

DNA	AMOUNT
Genomic	1 ng–1 μg
Plasmid or Viral	1 pg–1 ng

2. **Primers:**
Oligonucleotide primers are generally 20–40 nucleotides in length and ideally have a GC content of 40–60%. Computer programs such as Primer3 (<http://frodo.wi.mit.edu/primer3>) can be used to design or analyze primers. The final concentration of each primer in a PCR reaction may be 0.05–1 μM, typically 0.1–0.5 μM.
3. **Mg⁺⁺ and additives:**
Mg⁺⁺ concentration of 1.5–2.0 mM is optimal for most PCR products generated with Crimson *Taq* DNA Polymerase. The Mg-free buffer formulation along with supplemental MgCl₂ solution gives the user complete control over the final Mg⁺⁺ concentration in the reaction.

Amplification of some difficult targets, like GC-rich sequences, may be improved with additives, such as DMSO (8) or formamide (9).

4. **Deoxynucleotides:**
The final concentration of dNTPs is typically 200 μM of each deoxynucleotide.
5. **Crimson Taq DNA Polymerase Concentration:**
We generally recommend using Crimson *Taq* DNA Polymerase at a concentration of 25–50 units/ml (1.25–2.5 units/50 μl reaction). However, the optimal concentration of Crimson *Taq* DNA Polymerase may range from 5–50 units/ml (0.25–2.5 units/50 μl reaction) in specialized applications.

(see other side)

6. Denaturation:
An initial denaturation of 30 seconds at 95°C is sufficient for most amplicons from pure DNA templates. For difficult templates such as GC-rich sequences, a longer denaturation of 2–4 minutes at 95°C is recommended prior to PCR cycling to fully denature the template. With colony PCR, an initial 5 minute denaturation at 95°C is recommended.

During thermocycling a 15–30 second denaturation at 95°C is recommended.

7. Annealing:
The annealing step is typically 15–60 seconds. Annealing temperature is based on the T_m of the primer pair and is typically 45–68°C. Annealing temperatures can be optimized by doing a temperature gradient PCR starting 5°C below the calculated T_m .

When primers with annealing temperatures above 60°C are used, a 2-step PCR protocol is possible (see #10).

8. Extension:
The recommended extension temperature is 68°C. Extension times are generally 1 minute per kb. A final extension of 5 minutes at 68°C is recommended.

9. Cycle number:
Generally, 25–35 cycles yields sufficient product. Up to 45 cycles may be required to detect low-copy-number targets.

10. 2-step PCR:
When primers with annealing temperatures above 60°C are used, a 2-step thermocycling protocol is possible.

Thermocycling Conditions for a Routine 2-Step PCR:

STEP	TEMP	TIME
Initial Denaturation	95°C	30 seconds
30 Cycles	95°C	15–30 seconds
	60–68°C	1 minute/kb
Final Extension	60–68°C	5 minutes
Hold	4–10°C	

11. PCR product:
The PCR products generated using Crimson *Taq* DNA Polymerase contain dA overhangs at the 3'-end; therefore the PCR products can be ligated to dT/dU-overhang vectors.

FAQs:

1. *What are the advantages or disadvantages of Crimson Taq DNA Polymerase?*

The Crimson *Taq* Reaction Buffer formulation offers three convenient features. First, the 5X Crimson *Taq* Reaction Buffer contains a red dye, which serves as a visual indicator of homogeneous reaction setup. Second, the 5X Crimson *Taq* Reaction Buffer contains a density reagent, dextran, which allows direct loading of PCR products on a gel. Third, the trace amount of red dye in Crimson *Taq* Reaction Buffer works as a gel tracking dye (migrates at about 10 bp) during electrophoresis.

If the PCR products will be analyzed by absorbance or fluorescence excitation, acid red (λ_{max} = 510 nm) may interfere with the assay; therefore Standard *Taq* Reaction Buffer is recommended.

2. *Does the 5X Crimson Taq Reaction Buffer offer amplification efficiency similar to that of Standard Taq Reaction Buffer or ThermoPol Reaction Buffer?*

Yes.

3. *Can the PCR product be used directly in T/A cloning?*

Yes. PCR products can be directly ligated to dT/dU-overhang vectors and then transformed directly into host cells.

4. *How do I remove the dye and dextran from my PCR reactions using Crimson Taq Reaction Buffer?*

Spin columns for PCR clean-up can be used to remove both the dye and dextran.

5. *How long is the buffer stable in non-optimal storage conditions?*

The buffer is best stored at -20°C, but full activity is retained in buffer that is stored at 4°C for two weeks. Storage at room temperature is not recommended for more than 4 days.

References:

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Companion Products Sold Separately:

Crimson *Taq* Reaction Buffer Pack
#B0324S 6.0 ml

Crimson *Taq* (Mg-free) Reaction Buffer Pack
#B0325S 6.0 ml

Diluent F
#B8006S 4.0 ml

Magnesium Chloride (MgCl₂) Solution
#B9021S 6.0 ml

Taq PCR Kit
#E5000S 200 Reactions

Taq PCR Kit with Controls
#E5100S 200 Reactions

Taq 2X Master Mix
#M0270S 100 Reactions
#M0270L 500 Reactions

Quick-Load® *Taq* 2X Master Mix
#M0271S 100 Reactions
#M0271L 500 Reactions

Taq 5X Master Mix
#M0285S 100 Reactions
#M0285L 500 Reactions

Crimson *Taq* DNA Polymerase
#M0324S 200 units
#M0324L 1,000 units

Deoxynucleotide Solution Set
#N0446S 25 µmol each

Deoxynucleotide Solution Mix
#N0447S 8 µmol each
#N0447L 40 µmol each