

#### PRODUCT INFORMATION

# **T4 RNA Ligase**

**#EL0021** 1000 u

Lot: Expiry Date:

Concentration: 10 u/µl

Supplied with: 0.2 ml of 10X Reaction Buffer

0.2 ml of 1 mg/ml BSA

Store at -20°C

BSA included

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#### **Description**

T4 RNA Ligase catalyzes the ATP-dependent intra- and intermolecular formation of phosphodiester bonds between 5'-phosphate and 3'-hydroxyl termini of oligonucleotides, single-stranded RNA and DNA. The minimal substrate is a nucleoside 3',5'-biphosphate in intermolecular reaction and oligonucleotide of 8 bases in intramolecular reaction.

## **Applications**

- RNA 3'-end labeling with cytidine 3',5'- bis [α-<sup>32</sup>P] phosphate (1).
- Joining RNA to RNA (2).
- Synthesis of oligoribonucleotides and oligodeoxyribonucleotides (3, 4).
- Specific modifications of tRNAs (5).
- Oligodeoxyribonucleotide ligation to single-stranded cDNAs for 5'-RACE (Rapid Amplification of cDNA Ends) (6).
- Site-specific generation of composite primers for PCR (7).

#### Source

E.coli cells with a cloned gene 63 of bacteriophage T4.

Rev.5



# **Definition of Activity Unit**

One unit of the enzyme catalyzes the conversion of 1 nmol of  $5'-[^{32}P]-(A)_{12-18}$  to a phosphatase-resistant form in 30 min at  $37^{\circ}C$ .

Enzyme activity is assayed in the following mixture: 50 mM Tris-HCl (pH 7.5), 10 mM MgCl $_2$ , 10 mM DTT, 1 mM ATP, 10  $\mu$ M 5'-[ $^{32}$ P]-(A) $_{12\cdot18}$  (10  $\mu$ M in 5'-termini).

### **Storage Buffer**

The enzyme is supplied in: 20 mM Tris-HCl (pH 7.5), 1 mM DTT, 50 mM KCl, 0.1 mM EDTA, 0.03% (v/v) ELUGENT Detergent and 50% (v/v) glycerol.

#### **10X Reaction Buffer**

500 mM Tris-HCl (pH 7.5 at 25°C), 100 mM MgCl $_{2}$ , 100 mM DTT, 10 mM ATP.

## **Inhibition and Inactivation**

- Inhibitors: metal chelators, SH group-modifying reagents (8).
- Inactivated by heating at 70°C for 10 min.

#### Note

The recommended BSA concentration in the reaction mixture is 0.1 mg/ml.

#### **CERTIFICATE OF ANALYSIS**

# **Endodeoxyribonuclease Assay**

No conversion of covalently closed circular DNA to nicked DNA was detected after incubation of 50 units of T4 RNA Ligase with 1  $\mu$ g of pUC19 DNA for 4 hours at 37°C.

## **Ribonuclease Assay**

No contaminating RNase activity was detected after incubation of 50 units of T4 RNA Ligase with 1  $\mu$ g of [³H]-RNA for 4 hours at 37°C.

### **Labeled Oligonucleotide (LO) Assay**

No degradation of single-stranded and double-stranded labeled oligonucleotide was observed after incubation with 20 units of T4 RNA Ligase for 4 hours at 37°C.

**Quality authorized by:** 



#### **References**

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- 2. Middleton, T., et al., Synthesis and purification of oligoribonucleotides using T4 RNA ligase and reverse-phase chromatography, Anal. Biochem., 144, 110-117, 1985.
- 3. Brennan, C.A., et al., Using T4 RNA ligase with DNA substrates, Meth. Enzymol., 100, 38-52, 1983.
- 4. Tessier, D.C., et al., Ligation of single-stranded oligodeoxyribonucleotides by T4 RNA ligase, Anal. Biochem., 158, 171-178, 1986.
- Heckler, T.G., et al., T4 RNA ligase mediated preparation of novel "chemically misacylated" tRNA<sup>Phe</sup>s, Biochemistry, 23, 1468-1473, 1984.
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- 7. Kaluz, S., et al., Enzymatically produced composite primers: an application of T4 RNA ligase-coupled primers to PCR, BioTechniques, 19, 182-186, 1995.
- 8. Eun, H, M., Enzymology Primer for Recombinant DNA Technology, Academic Press. Inc., 1996.

#### **PRODUCT USE LIMITATION**

This product is developed, designed and sold exclusively *for research purposes and in vitro use only.* The product was not tested for use in diagnostics or for drug development, nor is it suitable for administration to humans or animals. Please refer to <a href="https://www.thermoscientific.com/fermentas">www.thermoscientific.com/fermentas</a> for Material Safety Data Sheet of the product.

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